



Phytoremediation Potential of *Jatropha gossypifolia* for Toxic metals in Waste-Impacted Soils Amended with Citric and Ethylenediaaminetetraacetic Acids

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Abstract: Phytoremediation is a recent technique that used plant for the extraction of toxic substances from impacted environment. The phytoextraction of Cd, Cr, Ni, and Pb from waste dumpsite soil (WDS), automobile waste-impacted soil (AIS), electronic waste-impacted soil (EWS), and paint waste-impacted soil (PIS) by the leaves and roots of *Jatropha gossypifolia* was assessed in this study. The seeds of *J. gossypifolia* were cultivated in forty-eight (48) bags in a set of twelve for each impacted soil amended with 5.0, 10.0, 15.0, and 20.0 mLday⁻¹ of citric acid (CA), ethylenediaaminetetraacetic acid (EDTA) and mixed CA+EDTA chelants and four (4) soils without amendment (one for each set of impacted soil) were used as the Controls. The cultivated seeds in polyethylene bags were watered daily for twelve (12) weeks. *J. gossypifolia* cultivated in the amended soils and those without amendment were harvested after 12 weeks, treated and analysed. The background impacted soils were treated and analysed for the concentrations of these toxic metals. The results revealed that the concentrations (mgkg⁻¹) of Cd, Cr, Ni, and Pb in the background soils varied as follows: 2.02-3.86, 5.09-7.71, 8.59-13.44, and 34.18-39.36, respectively. The mean concentrations of all the metals except Cd were within their acceptable limits. The concentrations of metals extracted by the roots of *J. gossypifolia* harvested from the studied and control soils were higher than the leaves. The concentrations of the metals in the amended soils varied with the concentrations in the background soils. Higher concentrations of toxic metals were extracted in the amended soils than the control. The potentials of the chelants in extracting the metals was in the order CA+EDTA > EDTA > CA. The rate of metal extraction by *J. gossypifolia* from the impacted soils was directly proportional to the concentration of the chelants. CA and mixed CA+EDTA improved the growth of *J. gossypifolia*, while EDTA inhibited the growth. The bioaccumulation factors (BCFs) of all the metals in the amended soils were higher than one, but less than one in the control soils. The translocation factors (TFs) of all the metals in the amended soils were higher than in the control pots. This study indicated that, *J. gossypifolia* has very high potential to extract toxic metals from impacted soils amended with chelants.

1. Introduction

Phytoremediation is a technique used for the reduction or elimination of toxic substances from impacted environments such as air, water, and land with suitable plants. This method is simple, economical, nature-based, eco-friendly, and improves the quality of soil (Asante-Badu *et al.*, 2020). Nevertheless, phytoremediation may be ineffective for contaminants such as polychlorinated biphenyls (PCBs). It can also cause contamination of the soil and underground water (Liu *et al.*, 2020). The process of phytoremediation is slow compared to the traditional excavation method of contaminated soils (Yan *et al.*, 2020). The various approaches involved in phytoremediation are (i) Phytoextraction: This involves the removal of contaminants in water or soil, and transferring them to the aboveground

parts of the plants (Jacob *et al.*, 2018; Suman *et al.*, 2018), (ii) Phytostabilization: This is a process whereby contaminants mostly metals are immobilized and accumulated by some metal-tolerant plants thereby reducing their availability within the environment (Lan *et al.*, 2020; Yan *et al.*, 2020), (iii) Phytovolatilization: This involves the use of plants to extract toxic metals from contaminated site, convert them into less toxic volatile form, and later release them into the atmosphere (Yan *et al.*, 2020; Kafle *et al.*, 2022), (iv) Phytofiltration: This is a process whereby plant roots (rhizofiltration), seedlings (blastofiltration), or shoots (caulofiltration) extract contaminants from impacted water or aqueous waste (Nedjimi, 2021), (v) Phytodegradation: This involves a practice whereby the roots of plant degrade organic contaminants in water or soil by the release of enzymes (Kafle *et al.*, 2022), and (vi) Rhizodegradation: This is the breaking down of organic contaminants in soil by microorganisms within the plant root zone or rhizosphere (Jha and Songachan, 2022; Chojnacka *et al.*, 2023). However, this study dealt exclusively on the phytoextraction and phytostabilization of toxic metals from impacted soils using *Jatropha gossypifolia* plant. *Jatropha gossypifolia* is plant that grows optimally in the tropics mostly in Nigeria within dumpsite soils (Awokunmi *et al.*, 2012; Mokhtari *et al.*, 2017; Awotedu *et al.*, 2018).

Toxic metals have adverse negative effects on human, animal, and plant lives hence; they should be eliminated from the environment by any possible but eco-friendly method (Rai *et al.*, 2019; Okon *et al.*, 2023; Etuk *et al.*, 2023). Some plants naturally have the potentials to extract high levels of contaminants from the impacted environment, while most plants have low potential for the extraction of contaminants. Consequently, chelating agents are introduced into the impacted area to accelerate the process of phytoremediation by these plants (Aghelan *et al.*, 2021; Kafle *et al.*, 2022). Chelating agents are both organic and synthetic; citric acid (CA) is an example of organic chelator, while ethylenediaminetetraacetic acid (EDTA) belongs to the synthetic class. The organic chelating agents are biodegradable hence; they are suitable for use in large quantities (Diarra *et al.*, 2021). However, due to the efficient nature of EDTA, it is the widely used chelating agent (Mousavi *et al.*, 2021). Consequently, EDTA persists in soil thus; it can contaminate the soil, plant, surface and underground water, and plants (Guo *et al.*, 2015).

Studies have shown that numerous plants have been employed for the different techniques of phytoremediation (Lone *et al.*, 2008; Antoniadis *et al.*, 2021; Bortoloti and Baron, 2022). Nevertheless, the perfect plant for phytoremediation should be able to accumulate and tolerate high levels of toxic contaminants, easy to cultivate, and high biomass (Shabani and Sayadi, 2012). Although, *Jatropha gossypifolia* has these properties it has not been intensively applied in the area of phytoremediation mostly within the study area. Thus, this study was undertaken to explore the potentials of *Jatropha gossypifolia* for the remediation of toxic metals from different impacted soils. The various impacted soils examined were amended with citric and ethylenediaminetetraacetic acids. The study also aimed at assessing relatively the potentials of CA and EDTA as chelating agents. The subsequent impacts of these chelating agents on the growth of *Jatropha gossypifolia* will be examined. The rate of accumulation of toxic metals from the different impacted soils by the roots and leaves of the plant will be evaluated.

2. Methodology

2.1. Experimental design

Plastic bags of 20 cm and 10 cm in diameter and height respectively were filled with 1.50 kg of the air-dried impacted soils. The seeds of *Jatropha gossypifolia* obtained from Makurdi in Benue State, Nigeria, were planted in all the bags and allowed to germinate. A total of 52 bags were used for this

experiment and the bags were separated into four groups of twelve (12) bags each. A set of 12 bags each for soil from municipal waste dumpsite (WDS), automobile waste-impacted soil (AIS), electronic waste-impacted soil (EWS), and paint waste-impacted soil (PIS) all obtained in Akwa Ibom State, Nigeria. Each group of 12 bags was subdivided into four sets and for each set, 5.0, 10.0, 15.0, and 20.0 mLday⁻¹ of citric acid (CA), ethylenediaminetetraacetic acid (EDTA) and mixed CA + EDTA chelants were added separately. For each of the groups of 12 pots of impacted soils, one bag was without chelants and was used as the control bag. The entire bags were watered daily with 500 mL of distilled water except the day prior to harvest. The growth of the plant in each bag was monitored daily by measuring the height of stems and width of the leaves for twelve (12) weeks. At the end of 12 weeks, the plants were harvested from all the pots and treated for further experiments (Waziri *et al.*, 2016; Njoku *et al.*, 2020; Waseem *et al.*, 2024).

2.2. Sample collection, treatment, and digestion

At the end of twelve (12) weeks, *Jatropha gossypifolia* were harvested with hand trowel to avoid loss of plant part (Figure 1). The plants were washed with distilled water to remove dirt, and separated into leaves and roots. The leaves and roots of plant separated were air dried for three weeks, then ground and sieved as reported by Ebong *et al.* (2023). The impacted soil samples were sun dried for 72 hours, ground and sieved. One gram (1 g) each of the plant tissue and soils were digested with 10 mL of 1:1 (w/v) HNO₃: H₂O in a beaker. The mixture was homogenized, the beaker covered with a watch glass and heated to 95 °C, then refluxed for 15 minutes. The mixture was allowed to cooled, and then 5 mL concentrated HNO₃ added and refluxed again for 30 minutes. The beaker was covered with a ribbed beaker and allowed to evaporate to 5 mL. When this step was completed, the mixture was cooled and 2 mL of distilled water and 3 mL of 30 % H₂O₂ were added.

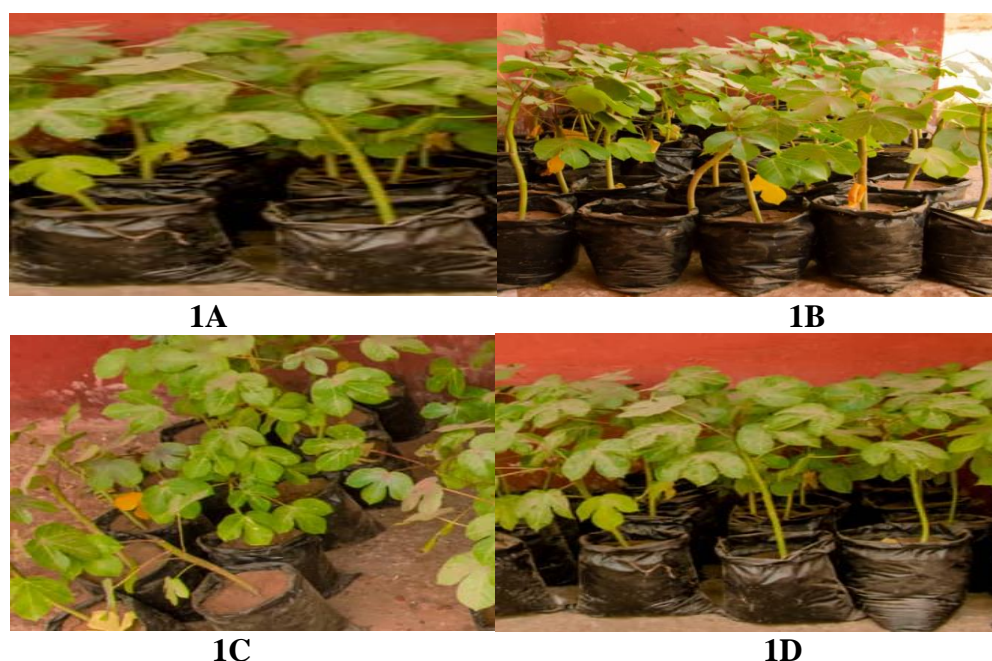


Figure 1. 1A= *J. gossypifolia* in control plot, 1B = *J. gossypifolia* in WDS + 20 mLday⁻¹ CA
1C = *J. gossypifolia* in WDS + 20 mLday⁻¹ EDTA, 1D = *J. gossypifolia* in WDS + 20 mLday⁻¹ CA+EDTA

The mixture was heated until effervescence from the peroxide reaction stopped. The mixture was cooled again and 1 mL of H₂O₂ added until effervescence was minimal. The mixture was cooled, 2 mL of concentrated HNO₃ and 10 mL distilled water added then the mixture was refluxed for 15 minutes

again. The mixture was centrifuged at 4000 rpm for 30 minutes to separate the filtrate from the residue (Saa-Aondo *et al.*, 2024). The concentrations (mgkg^{-1}) of Cd, Cr, Ni, and Pb in the filtrates were determined using Inductively Coupled Plasma Optical Emission spectroscopy (ICP-OES) (Shahbazi and Beheshti, 2019; Njoku and Nwani, 2022).

2.3. Determination of Bioaccumulation factor (BCF)

The bioaccumulation factor (BCF) of toxic metals in *Jatropha gossypifolia* was determined using Eqn. 1

$$\text{BCF} = \frac{C_{\text{mroot}}}{C_{\text{msoil}}} \quad \text{Eqn. 1}$$

where BCF is the bioaccumulation factor, C_{mroot} is the concentration of toxic metal in root, and C_{msoil} is the concentration of toxic metal in soil.

2.4. Determination of translocation factor (TF)

The translocation factor (TF) of toxic metals in *Jatropha gossypifolia* was calculated using Eqn. 2 below:

$$\text{TF} = \frac{C_{\text{mleaf}}}{C_{\text{mroot}}} \quad \text{Eqn. 2}$$

where TF is the transfer factor, C_{mleaf} represents the concentration of toxic metal in the leaf, C_{mroot} is concentration of toxic metal in the root.

2.5. Data Treatment

Data obtained in this study were subjected to statistical analysis using IBM SPSS Statistic version 29.0.2.0 (20) Software. The mean, maximum, minimum, and standard deviation (SD) were obtained directly from the Software. Multivariate analyses were performed by Varimax Factor analysis and Dendrograms using average linkage.

3. Results and Discussion

3.1. Concentrations of Toxic Metals in the Background Impacted Soils

The results of toxic metals in the studied impacted soils are shown in Table 1 above, cadmium (Cd) ranged from 2.02 to 3.86 mgkg^{-1} . The highest concentration of Cd was obtained in EWS, while the lowest was recorded for PIS. The reported high level of Cd in electronic waste-impacted soil is similar to the findings by Manikandan *et al.* (2023). The average value of Cd obtained ($3.00 \pm 0.82 \text{ mgkg}^{-1}$) is above the recommended limit for unpolluted soil by FAO/WHO (1999). Consequently, cultivation of edible crops may result in severe negative impacts on the consumers' health (Bouida *et al.*, 2022; Karim *et al.*, 2016). The concentrations of Cd at the different impacted soils examined followed the order EWS > WDS > ARS > PIS. This indicates that, the lowest concentration of Cd was recorded in the paint waste-impacted soil. Chromium (Cr) in the studied impacted soils ranged from 5.09 to 7.71 mgkg^{-1} between AIS and PIS, respectively. These results correspond with the one obtained by Woldeamanuale and Hassen, (2017). Thus, paint-related wastes have the potential of contaminating the environment with high level of Cr. However, the mean value of Cr obtained ($6.44 \pm 1.09 \text{ mgkg}^{-1}$) is within the safe limit of 100.0 mgkg^{-1} by FAO/WHO (1999). Nevertheless, as metals have the capacity to bio-accumulate and persist in an environment, it can still be a serious problem over time (Ali *et al.*, 2019; Ullah *et al.*, 2023). On the other hand, the levels of Cr obtained in this study could support plant

growth (Christou *et al.*, 2021; Gao *et al.*, 2022). The concentrations of Cr recorded in the different studied soils revealed the following trend PIS > WDS > EWS > ARS. Thus, automobile wastes showed lowest potential of contaminated the soil with Cr in the study.

Table 1: Concentration (mgkg⁻¹) of Toxic Metals in the Background Impacted soils

SAMPLE	Cd	Cr	Ni	Pb
WDS	3.45	6.73	11.23	39.36
AIS	2.67	5.09	13.44	35.14
EWS	3.86	6.23	12.31	35.70
PIS	2.02	7.71	8.59	34.18
MIN	2.02	5.09	8.59	34.18
MAX	3.86	7.71	13.44	39.36
MEAN	3.00	6.44	11.39	36.10
SD	0.82	1.09	2.08	2.27
ASL	0.80	100.0	35.0	85.0

*WDS = Waste dumpsite soil, AIS = Automobile waste-impacted soil, EWS = Electronic waste-impacted soil, PIS = Paint waste-impacted soil, MIN = Minimum, MAX = Maximum, SD = Standard deviation, ASL = Acceptable safe limit

The concentrations of nickel (Ni) in the soils examined varied from 8.59 mgkg⁻¹ at PIS to 13.44 mgkg⁻¹ in AIS. The reported high concentration of Ni in automobile waste-impacted soil is consistent with the results obtained by Liu *et al.* (2024). The average value of Ni recorded (11.39±2.08 mgkg⁻¹) is within the acceptable limit of 35.0 mgkg⁻¹ for an unpolluted soil by FAO/WHO (1999). Hence, the levels of Ni at all the studied soils could be considered as contaminant rather than pollutant. According to Shahzad *et al.* (2018), the level of Ni in the impacted soil assessed can promote plant growth and enzymatic activities. The concentrations of Ni in the studied soils followed the sequence ARS > EWS > WDS > PIS. Thus, wastes from paint industry had the lowest potential of impacting the soil with Ni.

The concentrations of lead (Pb) recorded varied from 34.18 to 39.36 mgkg⁻¹ (Table 1). The highest level was reported in the sample from WDS, while the lowest was obtained in PIS. The high concentration of Pb obtained in waste dumpsite soil is in agreement with the report by Agbeshie *et al.* (2020). The mean value of Pb obtained is within the acceptable limit of 85.0mgkg⁻¹ recommended by FAO/WHO (1999). Thus, the level of Pb at each location examined may not have adverse negative effect on environment and human health. Nevertheless, Pb is a highly poisonous metal without any beneficial impact thus; the level in the studied soils should be closely monitored to avoid toxicity and the related risks (Collin *et al.*, 2022). The concentrations of Pb at the different studied impacted soils followed the order WDS > EWS > ARS > PIS. The results revealed that, paint-related wastes showed the lowest degree of soil contamination by Pb.

3.2. Multivariate Analyses of Toxic Metals in the Impacted Soils

3.2.1. Principal Component Analysis of Toxic Metals

Principal component analysis (PCA) is a means of establishing the source(s) of contaminants in a studied ecosystem (Liu *et al.*, 2023). The outcome of PCA of toxic metals in the studied impacted soils indicated two key sources with Eigen value over one (1) and a cumulative variance of 91.7% (Table 2). Factor one (F1) with Eigen value 2.39 donated 59.7% to the total variance. F1 had significant positive loadings on Cd and Ni, moderate positive loading on Pb, and strong negative loading on Cr.

This may be the impact of electronic and automobile-related wastes (Manikandan *et al.*, 2023; Liu *et al.*, 2024). The second factor (F2) with Eigen value of 1.28, contributed 32.0% of the total variance. The factor (F2) had moderate positive loading on Cr and a significant positive loading on Pb (Table 2). This could be the negative influence of paint-related wastes and wastes at the municipal dumpsite (Woldeamanuale and Hassen, 2017; Agbeshie *et al.*, 2020).

Table 2: Results of Principal component analysis of Toxic metals in the Background Impacted Soils

	F1	F2
Cd	0.762	0.493
Cr	-0.838	0.524
Ni	0.945	-0.327
Pb	0.459	0.810
Eigen value	2.39	1.28
% Variance	59.7	32.0
% Cumulative	59.7	91.7

3.2.2. Hierarchical Cluster Analysis of Toxic Metals in the Impacted Soils

This work employed Hierarchical cluster analysis (HCA) for the identification of toxic metals with similar source as reported by Ebong *et al.* (2019) and Ebong *et al.* (2022). The HCA in Figure 2 shows three outstanding clusters namely: (a) The first Cluster links automobile waste-impacted soil (AIS) with electronic waste-impacted soil (EWS), (b) The second Cluster connects waste dumpsite soil (WDS) alone, and (c) The third Cluster links paint waste-impacted soil (PIS) only. This indicates that automobile and electronic wastes may have contributed common kind of metal contaminants to the soil environment, whereas metal contaminants contributed by wastes at municipal dumpsite and paint industry might have been peculiar to the source (Vesselinov *et al.*, 2018; Khaled-Khodja *et al.*, 2023).

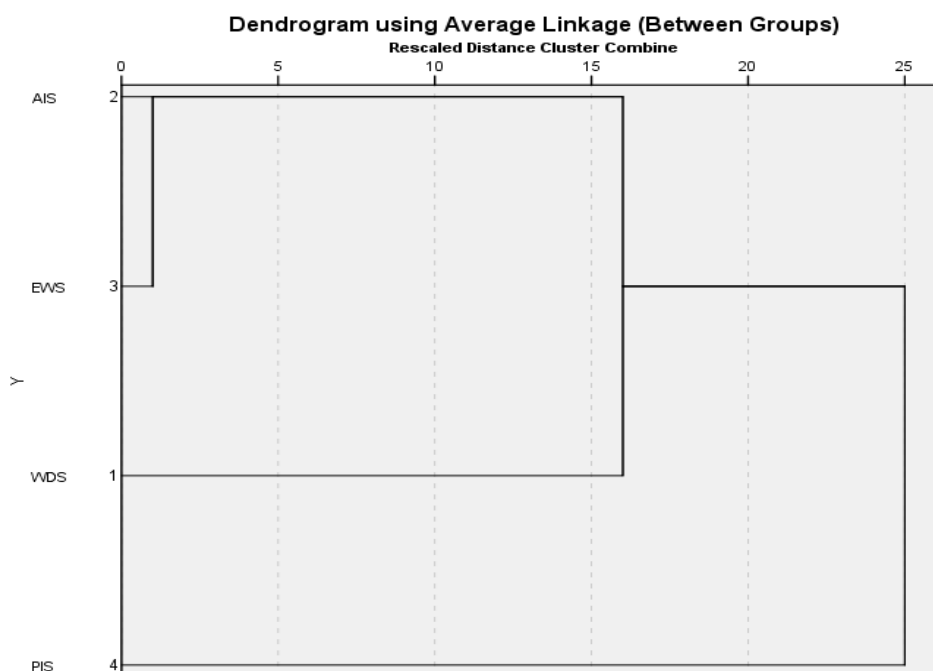


Figure 2: Hierarchical Clusters of Toxic Metals in the Impacted Soils

3.3. Concentrations of Toxic Metals Extracted by the Leaves and Roots of *J. gossypifolia* from the different Impacted Soils Amended with Chelants

The results of toxic metals extracted by the leaves of *J. gossypifolia* harvested from waste dumpsite soil (WDS) treated with different chelating agents are shown in **Table 3**. The concentrations (mgkg^{-1}) of toxic metals translocated to the leaves of *J. gossypifolia* harvested from WDS enhanced with various concentrations of citric acid (CA) as the chelating agent varied as follows: 0.07 – 0.11, 0.22 – 0.25, 0.38 – 0.42, and 1.55 – 1.59 for Cd, Cr, Ni, and Pb, respectively. Concentrations (mgkg^{-1}) of toxic metals translocated from WDS to the leaves of *J. gossypifolia* enhanced by different concentrations of EDTA were 0.08 – 0.12, 0.23 – 0.26, 0.39 – 0.43, and 1.56 – 1.60 for Cd, Cr, Ni, and Pb, respectively. Various concentrations of mixed CA+EDTA accelerated the translocation of 0.07 – 0.10, 0.18 – 0.21, 0.48 – 0.53, and 1.37 – 1.43 mgkg^{-1} of Cd, Cr, Ni, and Pb, respectively.

Table 3: Concentrations of Toxic Metals in the Leaves and Roots of *J. gossypifolia* from WDS Treated with Different Chelants

Conc. (mLday^{-1})	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	LEAVES				ROOTS			
CA								
5.0	0.07	0.22	0.38	1.55	0.08	0.24	0.41	1.57
10.0	0.09	0.23	0.40	1.57	0.10	0.25	0.42	1.58
15.0	0.10	0.24	0.41	1.58	0.11	0.26	0.42	1.59
20.0	0.11	0.25	0.42	1.59	0.12	0.29	0.43	1.61
EDTA								
5.0	0.08	0.23	0.39	1.56	0.09	0.25	0.42	1.58
10.0	0.10	0.24	0.41	1.58	0.11	0.26	0.43	1.59
15.0	0.11	0.25	0.42	1.59	0.12	0.27	0.44	1.60
20.0	0.12	0.26	0.43	1.60	0.13	0.30	0.45	1.62
CA + EDTA								
5.0	0.09	0.25	0.41	1.57	0.10	0.27	0.44	1.60
10.0	0.11	0.26	0.42	1.59	0.12	0.29	0.45	1.61
15.0	0.12	0.27	0.43	1.60	0.13	0.30	0.46	1.62
20.0	0.13	0.28	0.44	1.62	0.14	0.32	0.47	1.63

* WDS = WASTE DUMPSITE SOIL, CA = CITRIC ACID, EDTA = ETHYLENEDIAMINETETRAACETIC ACID

Table 3 shows the concentrations of toxic metals extracted from WDS by the roots of *J. gossypifolia* with various concentrations of CA chelant. The results varied for Cd, Cr, Ni, and Pb as follows: 0.08 – 0.12, 0.24 – 0.29, 0.41 – 0.43, and 1.57 – 1.61 mgkg^{-1} , respectively. The Concentrations of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* with EDTA as the chelating agent ranged as follows: 0.09 – 0.13, 0.25 – 0.30, 0.42 – 0.45, and 1.58 – 1.62 mgkg^{-1} , respectively. The concentrations (mgkg^{-1}) of toxic metals extracted by the roots of *J. gossypifolia* with mixed CA+EDTA at different concentrations as chelating agent varied respectively as 0.10 – 0.14, 0.27 – 0.32, 0.44 – 0.47, and 1.60 – 1.63 mgkg^{-1} for Cd, Cr, Ni, and Pb, respectively. The concentrations of all the toxic metals extracted by the roots of the studied plant from WDS were higher than the concentrations translocated to the leaves as reported by [Shehata et al. \(2019\)](#).

The results in **Table 3** indicate that, the leaves of *J. gossypifolia* harvested from waste dumpsite-impacted soil (WDS) extracted the following percentage of toxic metals 35.6, 44.4, 44.1, and

48.3 for Cd, Cr, Ni, and Pb, respectively. The proportions of Cd, Cr, Ni, and Pb extracted by the roots were 39.1, 49.1, 46.7, and 48.7%, respectively. The results reported confirmed that higher proportions of the metals were extracted by the roots than the leaves. This is consistent with the reports published by Riza and Hoque (2021) and Hemani *et al.* (2024) from similar studies. Generally, the leaves and roots of *J. gossypifolia* extracted a total of 74.7%, 93.5%, 90.8%, and 97.0% of Cd, Cr, Ni, and Pb from the contaminated soil (Table 7).

Table 4: Concentrations of Trace Metals in the Leaves and Roots of *J. gossypifolia* from AIS Treated with Different Chelants

Conc. (mLday ⁻¹)	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	LEAVES				ROOTS			
CA								
5.0	0.05	0.16	0.46	1.35	0.06	0.15	0.49	1.38
10.0	0.06	0.17	0.49	1.37	0.07	0.15	0.51	1.39
15.0	0.07	0.18	0.50	1.39	0.08	0.16	0.52	1.40
20.0	0.08	0.19	0.51	1.41	0.09	0.17	0.53	1.42
EDTA								
5.0	0.06	0.17	0.47	1.36	0.07	0.15	0.49	1.38
10.0	0.07	0.18	0.50	1.38	0.08	0.16	0.52	1.40
15.0	0.08	0.19	0.51	1.40	0.09	0.17	0.53	1.42
20.0	0.09	0.20	0.52	1.42	0.10	0.18	0.54	1.43
CA + EDTA								
5.0	0.07	0.18	0.48	1.37	0.08	0.16	0.51	1.42
10.0	0.08	0.19	0.50	1.40	0.09	0.17	0.54	1.43
15.0	0.09	0.20	0.52	1.41	0.10	0.18	0.55	1.44
20.0	0.10	0.21	0.53	1.43	0.11	0.20	0.56	1.46

* AIS = AUTOMOBILE WASTE-IMPACTED SOIL, CA = CITRIC ACID, EDTA = ETHYLENEDIAMINETETRA ACETIC ACID

The ranges of Cd, Cr, Ni, and Pb extracted from AIS by the leaves of *J. gossypifolia* with various concentrations of citric acid (CA) as the chelant were 0.05 – 0.08, 0.16 – 0.19, 0.46 – 0.51, and 1.35 – 1.41 mgkg⁻¹, respectively (Table 4). The applications of various concentrations of EDTA as chelating agent to AIS improved the concentrations of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* as indicated by the following ranges: 0.06 – 0.09, 0.17 – 0.20, 0.47 – 0.52, and 1.36 – 1.42 mgkg⁻¹, respectively (Table 4). The mixed CA+EDTA at various concentrations resulted in the extraction of 0.07 – 0.10 mgkg⁻¹ Cd, 0.18 – 0.21 mgkg⁻¹ Cr, 0.48 – 0.53 mgkg⁻¹ Ni, and 1.37 – 1.43 mgkg⁻¹Pb by the leaves of *J. gossypifolia* from AIS.

The concentrations (mgkg⁻¹) of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from AIS with different concentrations of CA chelant ranged as follows: 0.06 – 0.09, 0.15 – 0.17, 0.49 – 0.53 and 1.38 – 1.42 mgkg⁻¹, respectively (Table 4). The addition of EDTA at various concentrations enhanced the extraction of higher ranges (mgkg⁻¹) of Cd, Cr, Ni, and Pb by the roots of *J. gossypifolia* from AIS as follows: 0.07 – 0.10, 0.15 – 0.18, 0.49 – 0.54 and 1.38 – 1.43mgkg⁻¹, respectively (Table 4). Table 4 indicates the ranges of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from AIS after the addition of various concentrations of mixed CA+EDTA chelant as 0.08 – 0.11, 0.16 – 0.20, 0.51 – 0.56 and 1.42 – 1.46 mgkg⁻¹, respectively. Consequently, the concentrations of metals

extracted by both the leaves and roots of *J. gossypifolia* from AIS followed the order Pb > Ni > Cr > Cd as also reported for WDS. The results obtained from AIS revealed that, the concentrations of Cr translocated to the leaves of *J. gossypifolia* were higher than those extracted by the roots. This is similar to the results obtained by Mng'ong'o *et al.* (2021) from a related study. However, the concentrations of Cd, Ni, and Pb extracted by the roots were higher those obtained in the leaves. As shown in **Table 7**, the leaves of *J. gossypifolia* extracted 33.6, 43.6, 44.6, and 47.5% of Cd, Cr, Ni, and Pb, respectively from AIS. The roots extracted 38.1, 39.4, 46.9, and 48.3% of Cd, Cr, Ni, and Pb. Hence, a total of 71.7, 83.0, 91.5, and 95.8% of Cd, Cr, Ni, and Pb, respectively were extracted by both the leaves and roots of *J. gossypifolia* from AIS.

Table 5: Concentrations of Toxic Metals in the Leaves and Roots of *J. gossypifolia* from EWS Treated with Different Chelants

Conc. (mLday ⁻¹)	LEAVES				ROOTS			
	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
CA								
5.0	0.09	0.18	0.42	1.42	0.10	0.20	0.44	1.38
10.0	0.10	0.20	0.44	1.44	0.11	0.21	0.45	1.39
15.0	0.12	0.21	0.45	1.45	0.12	0.22	0.47	1.41
20.0	0.13	0.22	0.46	1.47	0.14	0.24	0.49	1.43
EDTA								
5.0	0.11	0.19	0.43	1.43	0.12	0.22	0.46	1.40
10.0	0.11	0.21	0.45	1.45	0.13	0.23	0.47	1.41
15.0	0.12	0.22	0.46	1.46	0.14	0.24	0.49	1.42
20.0	0.14	0.23	0.48	1.48	0.15	0.25	0.50	1.44
CA + EDTA								
5.0	0.12	0.21	0.45	1.45	0.13	0.23	0.48	1.42
10.0	0.13	0.22	0.46	1.47	0.14	0.24	0.49	1.44
15.0	0.13	0.23	0.47	1.48	0.15	0.25	0.50	1.45
20.0	0.15	0.24	0.49	1.50	0.16	0.26	0.51	1.46

* EWS = AUTOMOBILE WASTE-IMPACTED SOIL, CA = CITRIC ACID, EDTA = ETHYLENEDIAMINETETRAACETICACID

Table 5 above shows the ranges of Cd, Cr, Ni, and Pb translocated to the leaves of *J. gossypifolia* from electronic waste-impacted soil (EWS) enhanced by CA chelant as 0.09 – 0.13, 0.18 – 0.22, 0.42 – 0.46, and 1.42 – 1.47 mgkg⁻¹, respectively between concentrations of 5.0 and 20.0 mLday⁻¹. The addition of EDTA chelant improved the ranges of Cd, Cr, Ni, and Pb translocated to the leaves of *J. gossypifolia* from EWS as 0.11 – 0.14, 0.19 – 0.23, 0.43 – 0.48, and 1.43 – 1.48 mgkg⁻¹, respectively between the concentrations of 5.0 and 20.0 mLday⁻¹. The ranges (mgkg⁻¹) of Cd, Cr, Ni, and Pb translocated to the leaves of *J. gossypifolia* with mixed CA+ EDTA were 0.12 – 0.15, 0.21 – 0.24, 0.45 – 0.49, and 1.45 – 1.50mgkg⁻¹ between 5.0 and 20.0 mLday⁻¹ CA+EDTA chelant.

The concentrations (mgkg⁻¹) of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from EWS were 0.10 – 0.14, 0.20 – 0.24, 0.44 – 0.49, and 1.38 – 1.43mgkg⁻¹ between 5.0 and 20.0 mLday⁻¹ concentrations of CA chelant. Ranges of 0.12 – 0.15, 0.22 – 0.25, 0.46 – 0.50, and 1.40 – 1.44 mgkg⁻¹ of Cd, Cr, Ni, and Pb were extracted by the roots of *J. gossypifolia* from EWS between the 5.0 and 20.0 mLday⁻¹ concentrations of EDTA. The ranges of Cd, Cr, Ni, and Pb extracted by the leaves of the studied plant were 0.13 – 0.16, 0.23 – 0.26, 0.48 – 0.51, and 1.42 – 1.46 mgkg⁻¹ between the mixed CA+EDTA chelant concentrations of 5.0 and 20.0mLday⁻¹. **Table 5** also indicates that higher concentrations of Pb were extracted by the leaves of *J. gossypifolia* than the roots, whereas the concentrations of Cd, Cr, and Ni were higher in the roots than leaves. This is consistent with the report

published by [Abdulkadir et al. \(2019\)](#) from a phytoremediation study. The proportions of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* from EWS were 37.5, 41.1, 44.4, and 49.0%, respectively ([Table 7](#)). The roots of the studied plant extracted 41.2, 44.8, 46.7, and 47.8% of Cd, Cr, Ni, and Pb, respectively. Thus, a total of 78.7, 85.9, 91.1, and 96.8% of Cd, Cr, Ni, and Pb were extracted from EWS by the leaves and roots of *J. gossypifolia* ([Table 7](#)).

Table 6: Concentrations of Toxic Metals in the Leaves and Roots of *J. gossypifolia* from PIS Treated with Different Chelants

Conc.(mLday ⁻¹)	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	LEAVES				ROOTS			
CA								
5.0	0.04	0.22	0.27	1.34	0.04	0.25	0.25	1.31
10.0	0.05	0.23	0.28	1.35	0.05	0.26	0.26	1.33
15.0	0.06	0.25	0.29	1.37	0.05	0.27	0.27	1.35
20.0	0.07	0.26	0.30	1.39	0.06	0.28	0.29	1.36
EDTA								
5.0	0.05	0.24	0.28	1.36	0.05	0.26	0.26	1.33
10.0	0.07	0.25	0.29	1.37	0.06	0.27	0.27	1.35
15.0	0.08	0.26	0.30	1.39	0.06	0.28	0.28	1.37
20.0	0.09	0.27	0.32	1.40	0.07	0.29	0.30	1.38
CA + EDTA								
5.0	0.06	0.25	0.30	1.38	0.06	0.27	0.27	1.34
10.0	0.07	0.26	0.32	1.39	0.07	0.28	0.29	1.36
15.0	0.09	0.27	0.33	1.40	0.07	0.29	0.29	1.38
20.0	0.10	0.28	0.35	1.42	0.08	0.31	0.31	1.39

* PIS = PAINT WASTE-IMPACTED SOIL, CA = CITRIC ACID, EDTA = ETHYLENEDIAMINETETRAACETIC ACID

The concentrations of toxic metals extracted by the leaves and roots of *J. gossypifolia* from paint waste-impacted soil (PIS) are indicated in [Table 6](#). The concentrations of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* from PIS amended with CA chelant varied as follows: 0.04 – 0.07, 0.22 – 0.26, 0.27 – 0.30, and 1.34 – 1.39 mgkg⁻¹, respectively between 5.0 and 20.0mLday⁻¹ concentrations. Concentrations of the metals extracted by the leaves of *J. gossypifolia* from PIS amended with various concentrations of EDTA chelant between 5.0 and 20.0mLday⁻¹ ranged as follows: 0.05 – 0.09, 0.24 – 0.27, 0.28 – 0.32, and 1.36 – 1.40 mgkg⁻¹ for Cd, Cr, Ni, and Pb, respectively. The use of different concentrations of mixed CA+EDTA for the amendment of PIS resulted in the extraction of 0.06 – 0.10, 0.25 – 0.28, 0.30 – 0.35, and 1.38 – 1.42 mgkg⁻¹ of Cd, Cr, Ni, and Pb, respectively.

[Table 6](#) also indicates the ranges of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from PIS with CA chelant as 0.04 – 0.06, 0.25 – 0.28, 0.25 – 0.29, and 1.31 – 1.36 mgkg⁻¹, respectively between the concentrations of 5.0 and 20.0mLday⁻¹. The ranges of 0.05 – 0.07, 0.26 – 0.29, 0.26 – 0.30, and 1.33 – 1.38 mgkg⁻¹ of Cd, Cr, Ni, and Pb, respectively were extracted from PIS amended with different concentrations of EDTA by the roots of the studied plant. Concentrations (mgkg⁻¹) of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from PIS amended with mixed CA+EDTA ranged as follows: 0.06 – 0.08, 0.27 – 0.31, 0.27 – 0.31, and 1.34 – 1.39mgkg⁻¹ between 5.0 and 20.0 mLday⁻¹ concentrations of the chelant. The results obtained at PIS revealed that, higher concentrations of Cd, Ni, and Pb were extracted by the leaves of *J. gossypifolia* than the roots nevertheless; higher concentrations of Cr were extracted by the roots than the leaves. This is in

agreement with the results published in literature (Sêkara *et al.*, 2005; Abdulkadir *et al.*, 2019; Coulibaly *et al.*, 2021). The quantities of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* from EWS were 41.11, 39.5, 42.3, and 48.6%, respectively. The roots of the studied plant extracted 35.7, 43.0, 38.9, and 47.6% of Cd, Cr, Ni, and Pb, respectively. Thus, a total of 76.8, 82.5, 81.2, and 96.2% of Cd, Cr, Ni, and Pb were extracted from EWS by the leaves and roots of *J. gossypifolia* (Table 7).

3.4. Proportions of Toxic Metals in the Leaves and Roots of *J. gossypifolia* harvested from the control soils

The results in Table 7 indicate that, the highest proportion of Cd was obtained in electronic waste-impacted soil (EWS), while the lowest was recorded for the automobile waste-impacted soil (AIS). This confirms the findings by Chakraborty *et al.* (2022) and Ishchenko (2019) that, Cd is mainly released to the soil environment by e-wastes. The high concentration of Cd extracted from EWS could be attributed to the presence of Ni-Cd batteries as reported by Khan *et al.* (2017) and Kubier *et al.* (2019). The highest proportion of Cr was extracted from the municipal waste dumpsite soil (WDS), while the lowest was extracted in paint waste-impacted soil (PIS). This result corroborates the reports by previous authors that, municipal waste dumpsite soils have elevated levels of Cr (Afolagboye *et al.*, 2020; Agbeshie *et al.*, 2020; Sanga and Pius, 2024). This could be due to the presence of agricultural, electroplated metallic, timber, and leather-related wastes at the dumpsite (Xia *et al.*, 2020). The highest quantity of Ni was extracted from the automobile waste-impacted soil (AIS), whereas the lowest Ni fraction was extracted from PIS. This is in agreement with the findings by the previous authors that, activities at the automobile mechanic workshops increase the concentrations of Ni in the host and adjoining soil environments (Vincent *et al.*, 2022; Iwegbue *et al.*, 2024).

Table 7: Total percentage of Toxic Metals extracted by the Leaves and Roots of *J. Gossypifolia* harvested from the various impacted soils investigated

SAMPLE		Cd	Cr	Ni	Pb
WDS	LEAVES	35.6	44.4	44.1	48.3
	ROOTS	39.1	49.1	46.7	48.7
	TOTAL	74.7	93.5	90.8	97.0
ARS	LEAVES	33.6	43.6	44.6	47.5
	ROOTS	38.1	39.4	46.9	48.3
	TOTAL	71.7	83.0	91.5	95.8
EWS	LEAVES	37.5	41.1	44.4	49.0
	ROOTS	41.2	44.8	46.7	47.8
	TOTAL	78.7	85.9	91.1	96.8
PIS	LEAVES	41.1	39.5	42.3	48.6
	ROOTS	35.7	43.0	38.9	47.6
	TOTAL	76.8	82.5	81.2	96.2

The high proportion of Ni extracted from AIS could be attributed mostly to the combustion of petrol and diesel at these workshops (Shahzad *et al.*, 2018; Bartzas *et al.*, 2021). The highest quantity of Pb was extracted from WDS, while the lowest proportion was extracted from AIS. The high proportion of Pb extracted from WDS is consistent with the previous studies documented in literature (Akanchise *et al.*, 2020; Ibrahim *et al.*, 2020; Onwukeme and Eze, 2021). The high quantity of Pb extracted from WDS could be attributed to the presence of domestic, metallic, plastic, and agricultural wastes and batteries at the studied dumpsite (Twumasi *et al.*, 2016; Zhou *et al.*, 2022). The variations in the quantity of toxic metals extracted from one impacted soil to the other could be influenced mainly by

the difference in soil properties (Farrag *et al.*, 2011; Cui *et al.*, 2021; Wieczorek *et al.*, 2023). The following observations were deduced generally from the results obtained:

- (i) The proportions of the metals extracted were directly proportional to the concentrations of the metals in the impacted soils before the amendment with chelants as reported by Orr *et al.* (2020) and Alawsy *et al.* (2024).
- (ii) The potential of chelating agent used generally followed the trend CA+EDTA > EDTA > CA hence; the highest concentrations in all the studied impacted soils were extracted by mixed chelants, while CA extracted the lowest concentrations of the metals. This is in agreement with the findings by Guo *et al.* (2018) and Yang *et al.* (2022). The reported trend also revealed that, EDTA chelant was more effective in the phytoremediation study than CA (Shinta *et al.*, (2021); Nawaza *et al.*, 2022).
- (iii) The extraction of metals based on their concentrations followed the increasing order 5.0 > 10.0 > 15.0 > 20.0 mL day⁻¹. Accordingly, the concentrations of metals extracted varied directly with the concentration of the chelants as reported by Olusegun and Oluwafemi, (2012); Boulouizet *al.*, (2020); Khare *et al.* (2024); Tayiem *et al.* (2025).
- (iv) The concentrations of the toxic metals extracted from all the impacted soils examined followed the trend Pb > Ni > Cr > Cd. Hence, the highest metal extracted by *J. gossypifolia* was Pb, while Cd was the lowest. The high concentrations of Pb extracted by *J. gossypifolia* reported is similar to the results reported in literature (Gzar *et al.*, 2014; Li and Gao, 2019; Tatu *et al.*, 2020). Njoku and Nwani (2022) also documented from their phytoremediation study that, the lowest metal extracted from impacted soils was recorded for Cd.

3.5. Results of Toxic Metals extracted by the Leaves and Roots of *J. gossypifolia* harvested from the Control Soils

Results for the concentrations of toxic metals extracted by the leaves and roots of *J. gossypifolia* harvested from the control pots are shown in Table 8. The concentrations (mg kg⁻¹) of toxic metals translocated from the municipal waste dumpsite soil (WDS) without amendment (Control) to the leaves of *J. gossypifolia* were 0.56, 0.78, 0.83, and 3.15 mg kg⁻¹ for Cd, Cr, Ni, and Pb, respectively. The concentrations of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from the control pot of WDS were 0.73, 0.96, 1.10, and 4.32 mg kg⁻¹, respectively. The proportions of the metals translocated from the control pot of WDS to the leaves of *J. gossypifolia* were 16.2, 11.6, 7.4, and 8.0% for Cd, Cr, Ni, and Pb, respectively. The roots extracted 21.2, 14.3, 9.8, and 11.0% of Cd, Cr, Ni, and Pb, respectively from the control pot of WDS. The trend for the concentrations of toxic metals extracted from the control pot of WDS by the leaves and roots of *J. gossypifolia* was Pb > Ni > Cr > Cd. Thus, the trend for both the leaves and roots were similar.

Table 8 shows that the concentrations of Cd, Cr, Ni, and Pb, translocated from the control pot of AIS to the leaves of *J. gossypifolia* were 0.40, 0.61, 0.53, and 2.19 mg kg⁻¹, respectively. The roots of the studied plant extracted 0.74, 0.83, 0.90, and 3.47 mg kg⁻¹ of Cd, Cr, Ni, and Pb, respectively from the control pot of AIS. The concentrations of metals extracted from the control soil of AIS followed the orders Pb > Cr > Ni > Cd and Pb > Ni > Cr > Cd, respectively for the leaves and roots of *J. gossypifolia*.

The proportion of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* from AIS not amended by chelants were 15.0, 12.0, 3.9, and 6.2%, respectively. The proportions of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from AIS not treated with chelants were 27.7, 16.3, 6.7, and 9.9%, respectively. The concentrations (mg kg⁻¹) of Cd, Cr, Ni, and Pb translocated from the roots to the leaves of *J. gossypifolia* control pot of EWS were 0.58, 0.47, 0.49, and 1.73 mg kg⁻¹, respectively.

This follows the sequence Pb > Cd > Ni > Cr. **Table 8** indicates that, the roots of *J. gossypifolia* extracted 0.97, 0.64, 0.62, and 2.68 mgkg⁻¹ of Cd, Cr, Ni, and Pb, respectively from the control pot of EWS. Thus, the concentrations of the toxic metals extracted by the roots followed the order Pb > Cd > Cr > Ni.

Table 8: Total concentrations of metals in leaves and roots of *J. gossypifolia* harvested from the different control soils

	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	LEAVES				ROOTS			
WDS	0.56	0.78	0.83	3.15	0.73	0.96	1.10	4.32
AIS	0.40	0.61	0.53	2.19	0.74	0.83	0.90	3.47
EWS	0.58	0.47	0.49	1.73	0.97	0.64	0.62	2.68
PIS	0.35	0.55	0.43	2.40	0.46	0.72	0.64	3.11

*WDS = Municipal wastes dumpsite soil, AIS = Automobile waste-impacted soil, EWS = Electronic waste-impacted soil, PIS = Paint waste-impacted soil

The proportions of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* from the control pot of EWS were 15.0, 7.5, 4.0, 4.9%, respectively. The fractions of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from the EWS not amended with chelants were 25.1, 10.3, 5.0, and 7.5%, respectively. The leaves of *J. gossypifolia* extracted 0.35, 0.55, 0.43, and 2.40 mgkg⁻¹ of Cd, Cr, Ni, and Pb, respectively from the control pot of PIS (**Table 8**). The concentrations (mgkg⁻¹) of Cd, Cr, Ni, and Pb extracted by the roots of *J. gossypifolia* from the control pot of AIS were 0.46, 0.72, 0.64, and 3.11 mgkg⁻¹, respectively. The results obtained for the concentrations of toxic metals extracted by both the leaves and roots of the studied plants followed the order Pb > Cr > Ni > Cd.

The amounts of Cd, Cr, Ni, and Pb extracted by the leaves of *J. gossypifolia* from the control pot of PIS were 17.3, 7.1, 5.0, and 7.0%, respectively. The roots of the studied plant extracted 22.8, 9.3, 7.5, and 9.1% of Cd, Cr, Ni, and Pb, respectively from the control pot of PIS. The results for the total proportions of toxic metals extracted by both the leaves and roots of *J. gossypifolia* from the control pots as illustrated in **Figure 3** were 37.4, 25.9, 17.2, and 19.0% for Cd, Cr, Ni, and Pb in the control pot of WDS. In the control pot of AIS, the leaves and roots of *J. gossypifolia* collectively extracted 42.7% Cd, 28.3% Cr, 10.6% Ni, and 16.1% Pb (**Figure 3**). The leaves and roots of *J. gossypifolia* extracted a total of 40.1, 17.8, 9.0, and 12.4% of Cd, Cr, Ni, and Pb, respectively from EWS (**Figure 3**). **Figure 3** shows that 40.1%, 16.4%, 12.5% and 16.1%, respectively of total proportions of Cd, Cr, Ni, and Pb were extracted by the leaves and roots of *J. gossypifolia* from the control pot of PIS.

The results obtained from the control pots revealed that, the concentrations of all the metals were higher in the roots of *J. gossypifolia* than the leaves. Consequently, the study has confirmed that, the application of chelants to a contaminated soil can enhance the translocation of metals to the shoots of the plant during phytoremediation (Chen *et al.*, 2010, Lateefat *et al.*, 2023). Generally, the proportions of the toxic metals extracted by both the leaves and roots of *J. gossypifolia* followed a decreasing order of Cd > Cr > Pb > Ni. Obviously, the proportions of toxic metals extracted from the impacted soils amended with chelants were much higher than in the control pot as reported by Shehata *et al.* (2019) and Helaoui *et al.* (2023). Accordingly, the application of chelants to the impacted soils promoted the extraction of toxic metals by *J. gossypifolia* (Bian *et al.*, 2018; Dong *et al.*, 2023; Lateefat *et al.*, 2023; Zulkernain *et al.*, 2023).

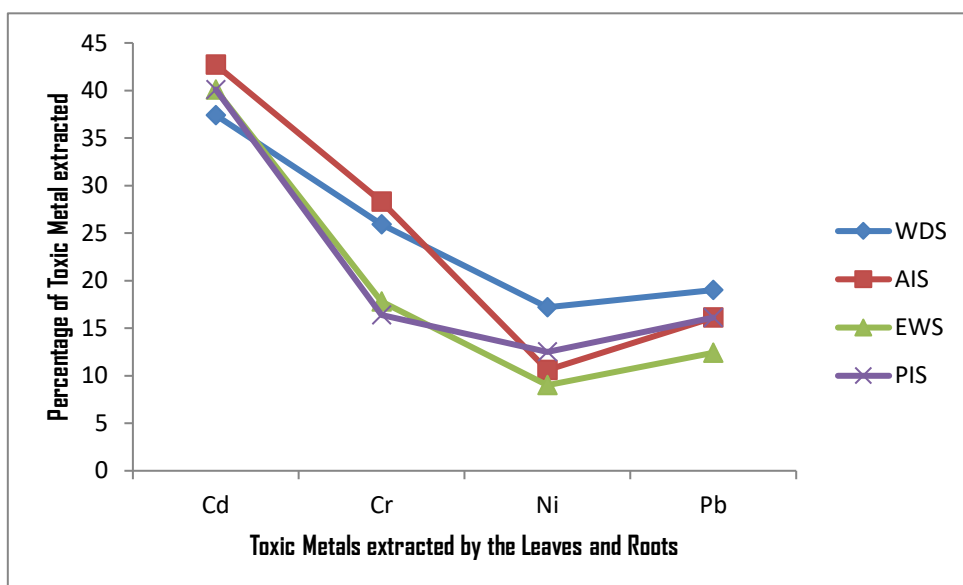


Figure 3: Total Percentage of Toxic Metals phytoextracted from the Control Pots

3.6. Results of Toxic Metals in the amended and controls after phytoextraction by *J. gossypifolia*

The results of the fractions of toxic metals in the control and amended soils after extraction by *J. gossypifolia* are shown in **Table 9**. The proportions (%) of Cd in the control soils were 62.6, 57.3, 59.9, and 59.9 for WDS, AIS, EWS, and PIS, respectively. The amounts of Cr in the control soils of WDS, AIS, EWS, and PIS were 74.1, 71.7, 82.2, and 83.6%, respectively. The fractions of Ni in the control soils after phytoextraction by *J. gossypifolia* were 82.8, 89.4, 91.0, and 87.5% in WDS, AIS, EWS, and PIS, respectively. Pb in the control soils after phytoextraction by the studied plant were 81.0, 83.9, 87.6, and 83.9% in WDS, AIS, EWS, and PIS, respectively.

Table 9: The proportions of Toxic metals in control and amended soils after phytoextraction by *J. gossypifolia*

	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	CONTROL SOIL				AMENDED SOIL			
WDS	62.6	74.1	82.8	81.0	25.3	6.5	9.2	3.0
AIS	57.3	71.7	89.4	83.9	28.3	17.0	8.5	4.2
EWS	59.9	82.2	91.0	87.6	21.3	14.1	8.9	3.2
PIS	59.9	83.6	87.5	83.9	23.2	17.5	18.8	3.8

*WDS = Municipal wastes dumpsite soil, AIS = Automobile waste-impacted soil, EWS = Electronic waste-impacted soil, PIS = Paint waste-impacted soil

The fractions of Cd in the soils amended with chelants after phytoextraction by *J. gossypifolia* were 25.3, 28.3, 21.3, and 23.2% in the WDS, AIS, EWS, and PIS, respectively (**Table 9**). The amounts of Cr in the amended WDS, AIS, EWS, and PIS after extraction by the studied plant were 6.5, 17.0, 14.1, and 17.5%, respectively (**Table 9**). The results in Table xx indicate that the fractions of Ni in the amended WDS, AIS, EWS, and PIS after phytoextraction were 9.2, 8.5, 8.9, and 18.8%, respectively. The proportions of Pb in the soils amended with chelants varied as follows: 3.0, 4.2, 3.2, and 3.8% respectively for WDS, AIS, EWS, and PIS. Accordingly, the proportions of toxic metals extracted from the soils amended with chelants were much higher than that of the control soils (Adjia *et al.*, 2008; Yang *et al.*, 2021). This corroborates the observations that, chelants increase the rate of metals uptake by plants (Aghelan *et al.*, 2021; Hosseinniaee *et al.*, 2023).

3.7. Results of Bioaccumulation factor (BCF) of Toxic Metals *J. gossypifolia*

The results of the bioaccumulation factor (BCF) of toxic metals in soils without amendment (Control) and the amended soils are shown in **Table 10**. The mean BCF of toxic metals in the control soils were as follows: Cd (0.64), Cr (0.35), Ni (0.20), and Pb (0.25). The BCF of all the toxic metals in the entire control soils were less than one (1). This agrees with the report by [Olayinka-Olagunju et al. \(2021\)](#) from a similar study. Consequently, *J. gossypifolia* could be effectively employed as excluder of these metals in the control soils ([Usman et al., 2019](#); [Zakari and Audu, 2021](#)). The reported BCF values of the toxic metals in the control soils followed a descending order of Cd > Cr > Pb > Ni.

Table 10: Bioaccumulation Factor (BCF) of toxic metals from the Soil to *J. gossypifolia*

	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	CONTROL SOIL				AMENDED SOIL			
WDS	0.58	0.40	0.27	0.30	1.14	1.42	1.38	1.46
AIS	0.71	0.44	0.18	0.26	1.10	1.22	1.38	1.44
EWS	0.65	0.28	0.14	0.20	1.20	1.31	1.38	1.45
PIS	0.63	0.26	0.21	0.25	1.13	1.25	1.20	1.44
Mean	0.64	0.35	0.20	0.25	1.14	1.30	1.34	1.45

*WDS = Municipal wastes dumpsite soil, ARS = Automobile waste-impacted soil, EWS = Electronicwaste-impacted soil, PIS = Paint waste-impacted soil

In the impacted soils amended with chelants, the mean BCF obtained for Cd, Cr, Ni, and Pb were 1.14, 1.30, 1.34, and 1.45, respectively. The BCF of all the metals in all the amended soils were higher than one. Hence, *J. gossypifolia* could be regarded as hyperaccumulator in the amended soils and could effectively be used for the remediation of soil impacted with these metals ([Usman et al., 2019](#)). The mean BCF values of toxic metals in the studied soils followed the descending order of Pb > Ni > Cr > Cd. Accordingly, the high BCF recorded for Pb is similar to the results reported by [Anani and Olomukoro \(2020\)](#) and [Ombugadu et al. \(2023\)](#). It was also deduced from the results that, higher BCF were reported in the amended soils than the control soils ([Padmavathiamma and Li, 2010](#); [Sulaiman and Hamzah, 2018](#); [Embrandiri et al., 2017](#)). Hence, soil amendment by the application of chelants can increase the rate of metal accumulation by plants from a contaminated site ([Lateefat et al., 2023](#); [Zulkernain et al., 2023](#)).

3.8. Results of Translocation factor (TF) of Toxic Metals *J. gossypifolia*

The results of the translocation factor (TF) of toxic metals from the roots of *J. gossypifolia* to the leaves in both the amended soils and soils without amendment (Control) are shown in **Table 11**. TF is the ratio of a particular metal in the leaves of a plant to that of the roots ([Sanjosé et al., 2022](#)). The TF of Cd in the control's soils ranged between 0.54 in AIS and 0.77 in WDS. TF of Cr in control soils varied from 0.73 in EWS and 0.81 in WDS. In the control pots, the TF of Ni ranged from 0.59 in AIS to 0.79 in EWS. The TF of Pb in the control soils ranged between 0.63 in AIS and 0.77 in PIS. The TF of all the toxic metals in soils without amendment were less than one. As a result of this, *J. gossypifolia* could be effectively employed for the phytostabilization of these metals in the control soils ([Madanan et al., 2021](#); [Mocek-Płóćiniak et al., 2023](#)). The TF of toxic metals in the amended soils ranged from 0.88 to 1.15 between AIS and PIS for Cd. The TF of Cr in the amended soils varied between 0.90 in WDS and 1.11 in AIS. A range of 0.95 – 1.09 was recorded for Ni between WDS and PIS. The TF of Pb in the amended soils varied from 0.98 in AIS to 1.03 in EWS. The TF of all the toxic metals in WDS were less than one (**Table 11**).

Table 11: Translocation factor of toxic metals from the roots to the leaves of *J. gossypifolia* in the control and amended soils

	Cd	Cr	Ni	Pb	Cd	Cr	Ni	Pb
	CONTROL SOIL				AMENDED SOIL			
WDS	0.77	0.81	0.75	0.73	0.91	0.90	0.95	0.99
AIS	0.54	0.74	0.59	0.63	0.88	1.11	0.95	0.98
EWS	0.60	0.73	0.79	0.65	0.91	0.92	0.96	1.03
PIS	0.76	0.76	0.67	0.77	1.15	0.92	1.09	1.02
Mean	0.67	0.76	0.70	0.70	0.96	0.96	0.99	1.01

*WDS = Municipal wastes dumpsite soil, ARS = Automobile waste-impacted soil, EWS = Electronic waste-impacted soil, PIS = Paint waste-impacted soil

In AIS, the TF of Cr was higher than one but less than one for other metals. The TF of all the metals except Pb were less than one, while the TF of Cd and Ni, and Pb were higher than one but were less than one for Cr. Thus, *J. gossypifolia* could be effectively utilized for the phytoextraction of Cd, Cr, Ni, and Pb in paint, automobile, and electronic waste-impacted soils (Sajad *et al.*, 2020). The results also revealed that Cd, Cr, Ni, and Pb were highly mobile in the paint, automobile, and electronic waste-impacted soils (Nahar and Hossen, 2020). However, in the locations where the TF were less than one, *J. gossypifolia* could be used for phytostabilization of the metals in those locations (Eribo *et al.*, 2021; Naz *et al.*, 2024; Aziz *et al.*, 2023). The translocation factors of toxic metals were higher in the amended soils than the soils without amendment (Control). This agrees with the results obtained from previous phytoremediation studies by Embrandiri *et al.* (2017) and Kulsoom *et al.* (2024). Hence, the application of chelants can improve the translocation of metals from contaminated soils to the plant (Saifullah *et al.*, 2009; Zulkernain *et al.*, 2023).

3.9. Impacts of soil amendment with chelants on the growth of *J. gossypifolia*

The application of citric acid (CA) to the impacted soils enhanced the growth of *J. gossypifolia* (Figure 1B). The use of CA for the amendment of the studied soils improved the height of the plant and enlarged the leaves (Shareef *et al.*, 2022; Ali *et al.*, 2024). The results obtained indicated that, the growth of *J. gossypifolia* was directly proportional to the concentration of CA chelant (Farid *et al.*, 2017; Kaur *et al.*, 2017; Kalousek *et al.*, 2024). The results also indicated improved plant growth for the soils amended with CA than plant in the control soil as reported by Twaij *et al.* (2019).

As shown in Figure 1C, the application of EDTA reduced the growth of *J. gossypifolia* significantly, and the growth was inversely proportional to the concentration of EDTA (Wirosoedarmo *et al.*, 2018; Huang *et al.*, 2021; Hosseinniaee *et al.*, 2023). It was deduced that, while the application of CA promoted the growth of *J. gossypifolia*, EDTA inhibited the growth of the plants. This conforms to the results documented from previous studies (Han *et al.*, 2018; Hosseini *et al.*, 2021; Shinta *et al.*, 2021). Figure 1D indicates that, the addition of mixed CA+EDTA chelant promoted the growth of *J. gossypifolia* as previously reported by Saleem *et al.* (2020) and Nawaza *et al.* (2022). However, the rate of growth did not show a direct relationship with the concentration of mixed CA+EDTA.

Jatropha gossypifolia exhibited a steady growth in all the control pots with the number of weeks. However, the tallest height of the stem and largest width of the leaves were recorded for the plant cultivated in the control pot of WDS (Figure 1A). The *J. gossypifolia* cultivated in electronic waste-impacted soil (EWS) was the shortest. The luxuriant growth exhibited by *J. gossypifolia* in the control soil of WDS could be attributed to the high plant nutrients available at dumpsite soil (Obianefo

et al., 2017; Ogbuehi *et al.*, 2021). The growth of *J. gossypifolia* in the control soils without amendment followed a decreasing order of :

WDS > PIS > AIS > EWS.

Conclusion

Based on the results, it could be concluded that, *J. gossypifolia* could be applied for phyto remediation of soils impacted with toxic metals. Higher concentration of toxic substances could be extracted by plants from highly impacted soils. The study also revealed that EDTA is a more effective chelant than citric acid however; a mixture of the two has a higher potential as chelant. Higher concentrations of metals were extracted when higher concentrations of the chelants were applied. The application of EDTA can impact negatively on the growth of plants used for phyto remediation while; citric acid improves their growth. Hence, the use of EDTA as a chelant for soil amendment during phyto remediation study should be minimized. The application of chelants could also improve the uptake of toxic substances by plants.

References

- Abdulkadir, L. G., Aliero, A. A., Shehu, K., Muhammad, A. B. (2019) Phyto remediation Potential of Selected Plant Species on Lead Contaminated Soil. *Savanna Journal of Basic and Applied Sciences*, 1(2), 248-255. <http://www.sjbas.com.ng>
- Abdullah, N. A. F., Ang, L. S. (2018) Binding Sites of Deprotonated Citric Acid and Ethylenediaminetetraacetic Acid in the Chelation with Ba²⁺, Y³⁺, and Zr⁴⁺ and Their Electronic Properties: a Density Functional Theory Study. *Acta Chim. Slov.*, 65, 231–238.
- Adjia, R., Fezeu, W. M. L., Tchatchueng, J. B., Sorho, S., Echevarria, G., Ngassoum, M. B. (2008) Long term effect of municipal solid waste amendment on soil heavy metal content of sites used for periurban agriculture in Ngaoundere, Cameroon. *African Journal of Environmental Science and Technology*, 2(12), 412-421.
- Afolagboye, L. O., Ojo, A. A., Talabi, A. O. (2020) Evaluation of soil contamination status around a municipal waste dumpsite using contamination indices, soil-quality guidelines, and multivariate statistical analysis. *SN Appl. Sci.*, 2, 1864. <https://doi.org/10.1007/s42452-020-03678-y>
- Aghelan, N., Sobhanardakani, S., Cheraghi, M., Lorestani, B., Merrikhpour, H. (2021) Evaluation of some chelating agents on phyto remediation efficiency of *Amaranthus caudatus* L. and *Tagetes patula* L. in soils contaminated with lead. *J Environ Health Sci Eng.*, 19(1), 503-514. <https://doi.org/10.1007/s40201-021-00623-y>
- Agbeshie, A. A., Adjei, R., Anokye, J., Banunle, A. (2020). Municipal waste dumpsite: Impact on soil properties and heavy metal concentrations, Sunyani, Ghana. *Scientific African*, 8, e00390. <https://doi.org/10.1016/j.sciaf.2020.e00390>
- Aghelan, N., Sobhanardakani, S., Cheraghi, M., et al. (2021) Evaluation of some chelating agents on phyto remediation efficiency of *Amaranthus caudatus* L. and *Tagetes patula* L. in soils contaminated with lead. *J Environ Health Sci Eng.*, 19(1), 503-514. <https://doi.org/10.1007/s40201-021-00623-y>
- Akanchise, T., Boakye, S., Borquaye, L. S., Dodd, M., Godfred Darko, G. (2020) Distribution of heavy metals in soils from abandoned dump sites in Kumasi, Ghana. *Scientific African*, 10, e00614. <https://doi.org/10.1016/j.sciaf.2020.e00614>
- Alawsy, W. S. A., Alabadi, L. A. S., AL-jibury, D. A. (2024) Employing Phyto remediation Methods to Extract Heavy Metals from Polluted Soils. *Ecological Engineering & Environmental Technology*, 25(6), 352–361. <https://doi.org/10.12912/27197050/187775>
- Ali, H., Khan, E., Ilahi, I. (2019) Environmental Chemistry and Ecotoxicology of Hazardous Heavy Metals: Environmental Persistence, Toxicity, and Bioaccumulation. *Journal of Chemistry*, 2019: 6730305. <https://doi.org/10.1155/2019/6730305>
- Ali, N., Rafiq, R., Zaib-un-Nisa, Wijaya, L., et al. (2024) Exogenous citric acid improves growth and yield by concerted modulation of antioxidant defense system in brinjal (*Solanum melongena* L.) under salt-stress. *Journal of King Saud University - Science*, 36(1), 103012. <https://doi.org/10.1016/j.jksus.2023.103012>

- Anani, O. A., Olomukoro, J. O. (2020) Assessment of Metal Accumulation and Bioaccumulation Factor of Some Trace and Heavy Metals in Freshwater Prawn and Crab. *IntechOpen*. eBook (PDF) ISBN 978-1-83880-826-6, <https://doi.org/10.5772/intechopen.88103>
- Antoniadis, V., Shaheen, S. M., Stärk, H-J., Wennrich, R., Levizou, E., et al. (2021) Phytoremediation potential of twelve wild plant species for toxic elements in a contaminated soil. *Environment International*, 146, 106233. <https://doi.org/10.1016/j.envint.2020.106233>
- Asante-Badu, B., Kgorutla, L. E., Li, S. S., Danso, P. O., Xue, Z., Qiang, G. (2020) Phytoremediation of organic and inorganic compounds in a natural and agricultural environment: A review. *Applied Ecology and Environmental Research*, 18(5), 6875-6904. <https://doi.org/10.15666/aeer/180568756904>
- Awokunmi, E. E., Asaolu, S. S., Ajayi, O. O., Adebayo, O. A. (2012) "The Role of EDTA on Heavy Metals Phytoextraction by *Jatropha Gossypifolia* Grown on Soil Collected from Dumpsites in Ekiti State Nigeria". *International Journal of Environment and Climate Change*, 2(2), 153-162. <https://doi.org/10.9734/BJECC/2012/1291>
- Awotedu, O. L., Paul O. Ogunbamowo, P. O., Awotedu, B. F., Emmanuel, I-O, B. (2018) Comparative Growth Response of Three *Jatropha* Species on Heavy Metal Contaminated Soil. *International Annals of Science*, 5(1), 26-32.
- Aziz, R. A., Yiwen, M., Saleh, M., Salleh, M. N., Gopinath, S. C. B., Giap, S. G. E., Chinni, S. V., Gobinath, R. (2023) Bioaccumulation and Translocation of Heavy Metals in Paddy (*Oryza sativa* L.) and Soil in Different Land Use Practices. *Sustainability*, 15, 13426. <https://doi.org/10.3390/su151813426>
- Bartzas, G., Tsakiridis, P. E., Komnitsas, K. (2021) Nickel industry: Heavy metal(loid)s contamination - sources, environmental impacts and recent advances on waste valorization. *Current Opinion in Environmental Science & Health*, 21, 100253. <https://doi.org/10.1016/j.coesh.2021.100253>
- Bian, X., Cui, J., Tang, B., Li Yang, L. (2018) Chelant-Induced Phytoextraction of Heavy Metals from Contaminated Soils: A Review. *Pol. J. Environ. Stud.*, 27(6), 2417-2424. <https://doi.org/10.15244/pjoes/81207>
- Bortoloti, G.A., Baron D. (2022) Phytoremediation of toxic heavy metals by Brassica plants: A biochemical and physiological approach. *Environmental Advances*, 8, 100204. <https://doi.org/10.1016/j.envadv.2022.100204>
- Bouida, L., Rafatullah, M., Kerrouche, A., Qutob, M., Alosaimi, A. M., Alorfi, H. S., Hussein, M. A. (2022) A Review on Cadmium and Lead Contamination: Sources, Fate, Mechanism, Health Effects and Remediation Methods. *Water*, 14, 3432. <https://doi.org/10.3390/w14213432>
- Chakraborty, S. C., Qamruzzaman, M., Zaman, M. W., Alam, Md M., Hossain, Md D., Pramanik, B. K., Nguyen, L. N., Nghiem, L. D., Ahmed, M. F., Zhou, J. L., Md. Ibrahim. Mondal, H., Hossain, M. A., Johir, M. A. H., Ahmed, M. B., Sithi, J. A., Zargar, M., Moni, M. A. (2022) Metals in e-waste: Occurrence, fate, impacts and remediation technologies. *Process Safety and Environmental Protection*, 162, 230-252. <https://doi.org/10.1016/j.psep.2022.04.011>
- Chen, G., Li, J., Han, H., et al. (2022) Physiological and molecular mechanisms of plant responses to copper stress. *Int. J. Mol. Sci.*, 23(21), 12950. <https://doi.org/10.3390/ijms232112950>
- Chen, L., Liu, T., Ma, C. (2010) Metal complexation and biodegradation of EDTA and S,S-EDDS: a density functional theory study. *J Phys Chem A*, 114(1), 443-54. <https://doi.org/10.1021/jp904296m>
- Chojnacka, K., Moustakas, K., Mikulewicz, M. (2023) The combined rhizoremediation by a triad: plant-microorganism-functional materials. *Environ Sci Pollut Res Int.*, 30(39), 90500-90521. <https://doi.org/10.1007/s11356-023-28755-8>
- Christou, A., Georgiadou, E. C., Zissimos, A. M., Christoforou, I. C., Christofi, C., Neocleous, D., Dalias, P., Fotopoulos, V. (2021) Uptake of hexavalent chromium by *Lactuca sativa* and *Triticum aestivum* plants and mediated effects on their performance, linked with associated public health risks. *Chemosphere*, 267, 128912.
- Collin, S., Baskar, A., Geevarghese, D.M., Syed Ali, M.N.V.S., Bahubali, P., Choudhary R., Lvov V., Tovar G.I. et al. (2022) Bioaccumulation of lead (Pb) and its effects in plants: A review. *Journal of Hazardous Materials Letters*, 3, 100064. <https://doi.org/10.1016/j.hazl.2022.100064>
- Coulibaly, H., Ouattara, P., Messou, A., Coulibaly, L. (2021) Phytoextraction of Trace Metals (Cd, Ni and Pb) by *Panicum maximum* Grown on Natural Soil. *Open Journal of Applied Sciences*, 11, 929-945. <https://doi.org/10.4236/ojapps.2021.118068>
- Cui, X., Mao, P., Sun, S., Huang, R., Fan, Y., Li, Y., et al. (2021), Phytoremediation of cadmium contaminated soils by *Amaranthus Hypochondriacus* L.: The effects of soil properties highlighting cation exchange capacity. *Chemosphere*, 283, 131067. <https://doi.org/10.1016/j.chemosphere.2021.131067>

- Diarra, I., Kotra, K. K., Prasad, S. (2021) Assessment of biodegradable chelating agents in the phytoextraction of heavy metals from multi-metal contaminated soil. *Chemosphere*, 273, 128483. <https://doi.org/10.1016/j.chemosphere.2020.128483>
- Dong, W., Wang, R., Li, H., Yang, X., Li, J., Wang, H., Jiang, C., Wang, Z. (2023) Effects of Chelating Agents Addition on Ryegrass Extraction of Cadmium and Lead in Artificially Contaminated Soil. *Water*, 15, 1929. <https://doi.org/10.3390/w15101929>
- Ebong, G. A., Etuk, H. S. and Dan, E. U. (2019). Multivariate Statistical Evaluation of Ecological Risks Associated with the Uncontrolled Tipping Method of Urban Wastes at Uyo Village Road, Akwa Ibom State, Nigeria. *Singapore Journal of Scientific Research*, 9 (1), 1-12.
- Ebong, G. A., Etuk, H. S., Anweting, I. B., Ekot, A. E., Ite, A. E. (2023) Relationship between traffic density, metal accumulation, pollution status, and human health problems in roadside soils and vegetables within the South-South Region of Nigeria. *International Journal of Environment, Agriculture and Biotechnology*, 8(3), 65 -79. <https://doi.org/10.22161/ijeab.83.8>
- Ebong, G. A., Moses, E. A., Akpabio, O. A. and Udombeh, R. B. (2022) Physicochemical properties, total concentration, geochemical fractions, and health risks of trace metals in oil-bearing soils of Akwa Ibom State, Nigeria. *Journal of Materials & Environmental Sustainability Research*, 2(4), 1-18. <https://doi.org/10.55455/jmesr.2022.009>
- Embrandiri, A., Rupani, P.F., Shahadat, M., Singh, R. P., Ismail, S. A., Ibrahim, M. H., Abd.Kadir, M. O. (2017) The phytoextraction potential of selected vegetable plants from soil amended with oil palm decanter cake. *Int J Recycl Org Waste Agricult.*, 6, 37–45. <https://doi.org/10.1007/s40093-016-0150-6>
- Eribo, O., Oterai, S. O., Inegbenijesu, O. B. (2021) Bioconcentration and Translocation Factors of Heavy Metals in *Rhizophora racemosa* and Sediments from Egbokodo Mangrove Swamp, Delta State, Nigeria. *African Scientist*, 22(4), 135-140.
- Etuk, H. S., Ebong, G. A., Anweting, I. B., et al. (2023) Sources and health risks of inorganic toxicants in *Gallus gallus domesticus* (broilers) from Poultry Farms in Uyo, Nigeria. *British Journal of Multidisciplinary and Advanced Studies: Sciences*, 4(5), 1-30. <https://doi.org/10.37745/bjmas.2022.0302>
- FAO/WHO. (1999) Food and Agricultural Organization/World Health Organization (FAO/WHO). *Joint FAO/WHO Food Standards Programme, Codex Alimentarius Commission, Twenty-third Session, 28 June - 3 July 1999, Rome, Italy*. Rome:
- Farid, M., Ali, S., Rizwan, M., Ali, Q., Abbas, F., Bukhari, S.A.H., Saeed, R., Wu, L. (2017) Citric acid assisted phytoextraction of chromium by sunflower; morpho-physiological and biochemical alterations in plants. *Ecotoxicol. Environ. Saf.*, 145, 90–102. <https://doi.org/10.1016/j.ecoenv.2017.07.016>
- Farrag K., Senesi N., Soler-Rovira P., Brunetti G. (2011) Effects of Selected Soil Properties on Phytoremediation Applicability for Heavy-Metal-Contaminated Soils in the Apulia Region, Southern Italy. *Environmental Monitoring and Assessment*, 184(11), 6593-606. <https://doi.org/10.1007/s10661-011-2444-5>
- Gao, H., Yang, X., Wang, N., Sun, M., Xiao, Y., Peng, F. (2022) Effects of Different Carbon Types on the Growth and Chromium Accumulation of Peach Trees under Chromium Stress. *Agronomy*, 12(11), 2814. <https://doi.org/10.3390/agronomy12112814>
- Guo, X., Wei, Z., Wu, Q. (2015) Degradation and residue of EDTA used for soil repair in heavy metal-contaminated soil. *Nongye Gongcheng Xuebao/Transactions of the Chinese Society of Agricultural Engineering*, 31(7), 272-278. <https://doi.org/10.3969/j.issn.1002-6819.2015.07.038>
- Guo, X., Zhao, G., Zhang, G., He, Q., Wei, Z., Zheng, W., Qian, T., Wu, Q. (2018) Effect of mixed chelators of EDTA, GLDA, and citric acid on bioavailability of residual heavy metals in soils and soil properties. *Chemosphere*, 209, 776-782. <https://doi.org/10.1016/j.chemosphere.2018.06.144>
- Gzar H., Abdul-Hameed A., Yahya A. (2014) Extraction of Lead, Cadmium and Nickel from Contaminated Soil Using Acetic Acid. *Open Journal of Soil Science*, 4, 207-214. <https://doi.org/10.4236/ojss.2014.46023>
- Han, Y., Zhang, L., Gu, J., Zhao, J., Fu, J. (2018) Citric acid and EDTA on the growth, photosynthetic properties and heavy metal accumulation of *Iris halophila* Pall. cultivated in Pb mine tailings. *International Biodeterioration & Biodegradation*, 128, 15 – 21. <https://doi.org/10.1016/j.ibiod.2016.05.011>
- Helaoui, S., Boughattas, I., Mkhinini, M., Chebbi, L., Elkribi-Boukhris, S., Alphonse, V., et al. (2023) Biochar amendment alleviates heavy metal phytotoxicity of *Medicago sativa* grown in polymetallic contaminated soil: Evaluation of metal uptake, plant response and soil properties. *Plant Stress*, 10, 100212. <https://doi.org/10.1016/j.stress.2023.100212>
- Hemani, Y., Malde, T., Puri, Y., Walvekar, S., D'souza, S. (2024) Effect of Heavy Metal Phytoremediation on Phytochemical Fingerprint and Bioactivity of *Pistia stratiotes*: A Quest for Re-routing Disposal to

- Commercial Application. *Nature Environment and Pollution Technology*, 23(3), 1579-1588. <https://doi.org/10.46488/NEPT.2024.v23i03.027>
- Hosseini, S. S., Lakzian, A., Halajnia, A., Razavi, B. S. (2021) Optimization of EDTA and citric acid for risk assessment in the remediation of lead contaminated soil. *Rhizosphere*, 17, 100277. <https://doi.org/10.1016/j.rhisph.2020.100277>
- Hosseinniaee, S., Jafari, M., Tavili, A., Zare, S., Cappai, G. (2023) Chelate facilitated phytoextraction of Pb, Cd, and Zn from a lead-zinc mine contaminated soil by three accumulator plants. *Sci Rep.*, 13(1), 21185. <https://doi.org/10.1038/s41598-023-48666-5>
- Huang, R., Cui, X., Luo, X., Mao, P., et al. (2021) Effects of plant growth regulator and chelating agent on the phytoextraction of heavy metals by *Pfaffia glomerata* and on the soil microbial community. *Environmental Pollution*, 283, 117159. <https://doi.org/10.1016/j.envpol.2021.117159>
- Ibrahim, G., Nwaichi, E., Abu, G. (2020) Heavy Metals Contents of Municipal Solid Waste Dumpsites in Potiskum, Yobe State Nigeria. *Journal of Environmental Protection*, 11, 709-717. <https://doi.org/10.4236/jep.2020.119043>
- Ishchenko V. (2019) Heavy metals in municipal waste: the content and leaching ability by waste fraction. *J Environ SciHealth A Tox Hazard Subst Environ Eng.*, 54(14), 1448-1456. <https://doi.org/10.1080/10934529.2019.1655369>
- Iwegbue, C. M. A., Nnanna C. A., Ogwu I. F., et al. (2024) Concentrations, sources and exposure to metals in dust from automobile mechanic workshops in Nigeria. *Journal of Trace Elements and Minerals*, 10, 100186. <https://doi.org/10.1016/j.jtemin.2024.100186>
- Jacob, J. M., Karthik, C., Saratale, R. G., Kumar, S. S., Prabakar, D., Kadirvelu, K., Pugazhendhi, A. (2018) Biological approaches to tackle heavy metal pollution: A survey of literature. *J Environ Manage.* 217, 56-70. <https://doi.org/10.1016/j.jenvman.2018.03.077>
- Jha, S. S., Songachan, L. S. (2022) Mycorrhizoremediation: Understanding the science behind it and its future prospects. *Materials Today: Proceedings*, 51(8), 2431-2436. <https://doi.org/10.1016/j.matpr.2021.11.605>
- Kafle, A., Timilsina, A., Gautam, A., Adhikari, K., Bhattarai, A., Aryal, N. (2022) Phytoremediation: Mechanisms, plant selection and enhancement by natural and synthetic agents. *Environmental Advances*, 8, 100203. <https://doi.org/10.1016/j.envadv.2022.100203>
- Kalousek, P., Holátko, J., Schreiber, P., [Pluháček, T.](#), [Lónová, K. Š.](#), [Radziemska, M.](#), [Tarkowski, P.](#), [Vyhnánek, T.](#), [Hammerschmidt, T.](#), [Brtnický, M.](#) (2024) The effect of chelating agents on the Zn-phytoextraction potential of hemp and soil microbial activity. *Chem. Biol. Technol. Agric.* 11, 23. <https://doi.org/10.1186/s40538-024-00544-6>
- Karim S., Aouniti A., El hajjaji F., Taleb M., et al. (2016), Bioaccumulation of heavy metals in commercially important marine fishes (*Palaemon Serratus* and *Solea Vulgaris*) caught in the Mediterranean coast from the North East of Morocco, *Der Pharma Chemica*, 8(19), 515-523
- Kaur, R., Yadav P., Sharma, A., Thukral, A. K., Kumar, V., et al. (2017) Castasterone and citric acid treatment restores photosynthetic attributes in *Brassica juncea* L. under Cd (II) toxicity. *Ecotoxicol. Environ. Saf.*, 145, 466–475. <https://doi.org/10.1016/j.ecoenv.2017.07.067>
- Khaled-Khodja, S., Cheraitia, H., Rouibah, K., et al. (2023). Identification of the Contamination Sources by PCBs Using Multivariate Analyses: The Case Study of the Annaba Bay (Algeria) Basin. *Molecules*, 28: 6841. <https://doi.org/10.3390/molecules28196841>
- Khan, M. A., Khan, S., Khan, A., Alam, M. (2017) Soil contamination with cadmium, consequences and remediation using organic amendments. *Sci. Total Environ*, 601, 1591–1605.
- Khare, S., Singhal, A., Rallapalli, S., Mishra, A. (2024) Bio-chelate assisted leaching for enhanced heavy metal remediation in municipal solid waste compost. *Sci Rep.*, 14, 14238. <https://doi.org/10.1038/s41598-024-65280-1>
- Kubier, A., Wilkin, R. T., Pichler, T. (2019) Cadmium in soils and groundwater: A review. *ApplGeochem.*, 108, 1-16. <https://doi.org/10.1016/j.apgeochem.2019.104388>
- Kulsoom, A. M., Yadav, A. K., Kumar, M., et al. (2024) Accumulation and Translocation of Heavy Metals in *Hibiscus cannabinus* Grown in Tannery Sludge Amended Soil. *Nature Environment and Pollution Technology*, 23(2), 1133-1139. <https://doi.org/10.46488/NEPT.2024.v23i02.047>
- Lan, M. M., Liu, C., Liu, S. J., Qiu, R. L., Tang, Y. T. (2020) Phytostabilization of Cd and Pb in Highly Polluted Farmland Soils Using Ramie and Amendments. *Int. J. Environ. Res. Public Health*, 17, 1661. <https://doi.org/10.3390/ijerph17051661>

- Lateefat, D. Y., Samaila, M. B., Ibrahim, S. (2023) Chelate Assisted Phytoremediation of Chromium from Soil Irrigated with Municipal Wastewater using *Calotropis procera*. *ChemSearch Journal*, 14(2), 90 – 97.
- Li, Q., Gao, Y. (2019) Remediation of Cd-, Pb- and Cu-Contaminated Agricultural Soils by Phosphate Fertilization and Applying Biochar. *Pol. J. Environ. Stud.*, 28(4), 2697-2705. <https://doi.org/10.15244/pjoes/92527>
- Liu, L., Luo, D., Lu, Y., Huang, X., Liu, Y., Wei, L., Xiao, T., Wu, Q., Liu, G. (2022) Risk assessment of groundwater pollution during GLDA-assisted phytoremediation of Cd- and Pb-contaminated soil. *Ecological Indicators*, 139, 108913. <https://doi.org/10.1016/j.ecolind.2022.108913>
- Liu, T., Ni, S. Y., Wang, Z. (2024) Contamination and health risk assessment of heavy metals in soil surrounding an automobile industry factory in Jiaying, China. *Front. Environ. Sci.*, 12, 1362366. <https://doi.org/10.3389/fenvs.2024.1362366>
- Liu, S., Yang, B., Liang, Y., Xiao, Y., Fang, J. (2020) Prospect of phytoremediation combined with other approaches for remediation of heavy metal-polluted soils. *Environ Sci Pollut Res Int.*, 27(14), 16069-16085. <https://doi.org/10.1007/s11356-020-08282-6>
- Lone M. I., He Z.L., et al. (2008) Phytoremediation of heavy metal polluted soils and water: progresses and perspectives. *J. Zhejiang Univ. Sci. B.*, 9(3), 210-20. <https://doi.org/10.1631/jzus.B0710633>
- Madanan, M. T., Shah, I. K., Varghese, G. K., Kaushal, R. K. (2021) Application of Aztec Marigold (*Tagetes erecta* L.) for phytoremediation of heavy metal polluted lateritic soil. *Environmental Chemistry and Ecotoxicology*, 3, 17 – 22. <https://doi.org/10.1016/j.enceco.2020.10.007>
- Manikandan, S., Inbakandan, D., Valli Nachiyar, C., Namasivayam, S. K. R. (2023) Towards sustainable metal recovery from e-waste: A mini review. *Sustainable Chemistry for the Environment*, 2, 100001. <https://doi.org/10.1016/j.scenv.2023.100001>
- Mng'ong'o, M., Munishi, L. K., Ndakidemi, P. A., Blake, W., et al. (2021) Accumulation and bioconcentration of heavy metals in two phases from agricultural soil to plants in Usangu agroecosystem-Tanzania. *Heliyon*, 7(7), e07514. <https://doi.org/10.1016/j.heliyon.2021.e07514>
- Mokhtari O., Abdellaoui S., Hamdani I., Aouniti A., Touzani R. (2017), A review on environmental and agricultural aspects of *Jatropha curcas*, *Arab. J. Chem. Environ. Res.* 04, 92-106
- Mousavi, A., Pourakbar, L., Moghaddam, S. S., Popovic-Djordjevic, J. (2021) The effect of the exogenous application of EDTA and maleic acid on tolerance, phenolic compounds, and cadmium phytoremediation by okra (*Abelmoschus esculentus* L.) exposed to Cd stress. *Journal of Environmental Chemical Engineering*, 9(4), 105456. <https://doi.org/10.1016/j.jece.2021.105456>
- Mocek-Plóćiniak, A., Mencil, J., Zakrzewski, W., Roszkowski, S. (2023) Phytoremediation as an Effective Remedy for Removing Trace Elements from Ecosystems. *Plants (Basel)*, 12(8), 1653. <https://doi.org/10.3390/plants12081653>
- Nahar, N., Hossen, S. (2020) Influence of sewage sludge application on soil properties, carrot growth and heavy metal uptake. *Communications in Soil Science and Plant Analysis*, 52(1), 1 – 10. <https://doi.org/10.1080/00103624.2020.1836201>
- Naz, R., Khana, M. S., Hafeezb, A., Fazila, M., Khana, M. N., et al. (2024) Assessment of phytoremediation potential of native plant species naturally growing in heavy metal-polluted industrial soils. *Brazilian Journal of Biology*, 84, e264473. <https://doi.org/10.1590/1519-6984.26447>
- Nawaza, H., Alib, A., Saleem, M. H., Ameer, A., Hafeeze, A., Alharbif, K., Ezzatg, A., Khana, A., Jamili, M., Faridj, G. (2022) Comparative effectiveness of EDTA and citric acid assisted phytoremediation of Ni contaminated soil by using canola (*Brassica napus*). *Brazilian Journal of Biology*, 82, e261785. <https://doi.org/10.1590/1519-6984.261785>
- Nedjimi, B. (2021) Phytoremediation: a sustainable environmental technology for heavy metals decontamination. *SN Appl. Sci.*, 3, 286. <https://doi.org/10.1007/s42452-021-04301-4>
- Njoku, K. L., Nwani, S. O. (2022) Phytoremediation of heavy metals contaminated soil samples obtained from mechanic workshop and dumpsite using *Amaranthus spinosus*. *Scientific African*, 17, e01278. <https://doi.org/10.1016/j.sciaf.2022.e01278>
- Njoku, K. L., Ojo, O. E., Jolaoso, A. O. (2020) Growth and ability of *Senna alata* in phytoremediation of soil contamination with heavy metals. *Not. Sci. Biol.*, 12(2), 420–432.
- Obianefo, F. U., Agbagwa, I. O., Tanee, F. B. G. (2017) Physicochemical Characteristics of Soil from Selected Solid Waste Dump Sites in Port Harcourt, Rivers State, Nigeria. *J. Appl. Sci. Environ. Manage.*, 21(6), 1153-1156. <https://dx.doi.org/10.4314/jasem.v21i6.27>

- Ogbuehi, H. C., Onuh, M. O., Christo, I. E. (2021) Effect of dumpsite organic manure soil on growth, nutrient content, and yield of *Curcuma longa* L.(turmeric) in Owerri, Imo State. *GSC Advanced Research and Reviews*, 08(01), 140–148. <https://doi.org/10.30574/gscarr.2021.8.1.0147>
- Okon, A. O., Ebong, G. A., Tombere, V. P., Anweting, I. B., Etuk, H. S., Ambrose, I. (2023) Toxicity profile of metals in water, sediments and *Liza grandisquamis* from Iko River, South-South of Nigeria. *International Journal of Frontline Research in Pharma and Biosciences*, 02(02), 001–016. <https://doi.org/10.56355/ijfrpbs.2023.2.2.0020>
- Olayinka-Olagunju, J. O., Dosumu, A. A., Olatunji-Ojo, A. M. (2021) Bioaccumulation of Heavy Metals in Pelagic and Benthic Fishes of Ogbese River, Ondo State, South-Western Nigeria. *Water Air Soil Pollut.*, 232, 44. <https://doi.org/10.1007/s11270-021-04987-7>
- Olusegun, A. O., Oluwafemi, A. S. (2012) Evaluation of chelating agents for the removal of heavy metals from contaminated soil. *Global Journal of Bio-Science and Biotechnology*, 1(2), 152-156.
- Ombugadu, A., Ahmed, H. O., Goler, E. E., Abok, J. I., Daan, S. A., Dogo, K. S., Markus, M. (2023) Comparative Study on Bioaccumulation of Lead (Pb) in Some Birds and Plant Species in Rapidly Degrading Mining Sites in Three Selected Local Government Areas of Nasarawa State, Central Nigeria. *Biomed J Sci & Tech Res.*, 53(3)-2023.
- Onwukeme V.I., Eze V.C. (2021) Identification of heavy metals source within selected active dumpsites in south eastern Nigeria. *Environ Anal Health Toxicol.*, 36(2), e2021008. <https://doi.org/10.5620/eaht.2021008>
- Orr, R., Hocking, R. K., Pattison, A., Nelson, P. N. (2020) Extraction of metals from mildly acidic tropical soils: Interactions between chelating ligand, pH and soil type. *Chemosphere*, 248, 126060. <https://doi.org/10.1016/j.chemosphere.2020.126060>
- Padmavathamma, P. and Li, L. Y. (2010) Effect of Amendments on Phytoavailability and Fractionation of Copper and Zinc in a Contaminated Soil. *International Journal of Phytoremediation*, 12(7), 697-715. <https://doi.org/10.1080/15226510903353179>
- Rai, P. K., Lee, S. S., Zhang, M., Tsang, Y. F., Kim, K-H. (2019) Heavy metals in food crops: Health risks, fate, mechanisms, and management. *Environment International*, 125, 365 – 385. <https://doi.org/10.1016/j.envint.2019.01.067>
- Riza, M., Hoque, S. (2021) Phytoremediation of Copper and Zinc Contaminated Soil around Textile Industries using Bryophyllumpinnatum Plant. *Journal of Ecological Engineering*, 22(4), 88-97. <https://doi.org/10.12911/22998993/134035>
- Saa-Aondo, M., A. Arose, A., Kalu, K. M., Yelwa, J. M., et al. (2024) Efficiencies of Heavy Metal Hyperaccumulation Plants as Potential Land Remediators for Heavy Metal Polluted Soils. *Asian Journal of Current Research*, 9(1), 60-70. <https://doi.org/10.56557/AJOCR/2024/v9i18525>
- Saifullah, Meers, E., Qadir, M., de Caritat, P., et al. (2009) “EDTA-Assisted Pb Phytoextraction.” *Chemosphere*, 74(10), 1279-91. <https://doi.org/10.1016/j.chemosphere.2008.11.007>
- Sajad, M. A., Khan, M. S., Bahadur, S., Shuaib, M., Naeem, A., Zaman, W., Ali, H. (2020) Nickel phytoremediation potential of some plant species of the Lower Dir, Khyber Pakhtunkhwa, Pakistan. *Limnol. Rev.*, 20, 13–22.
- Sanjosé, I., Navarro-Roldán, F., Montero, Y., Ramírez-Acosta, S., Jiménez-Nieva, F. J., Infante-Izquierdo, M. D., Polo-Ávila, A. and Muñoz-Rodríguez, A. F. (2022) The Bioconcentration and the Translocation of Heavy Metals in Recently Consumed *Salicornia ramosissima* J. Woods in Highly Contaminated Estuary Marshes and Its Food Risk. *Diversity*, 14, 452. <https://doi.org/10.3390/d14060452>
- Sanga, V.F., Pius, C.F. (2024) Heavy metal contamination in soil and food crops and associated human health risks in the vicinity of Iringa Municipal dumpsite, Tanzania. *Discov Environ.*, 2, 104. <https://doi.org/10.1007/s44274-024-00137-y>
- Sêkara, A., Poniedzia³ek, M., Ciura, J., Jêdrszczyk, E. (2005) Cadmium and Lead Accumulation and Distribution in the Organs of Nine Crops: Implications for Phytoremediation. *Polish Journal of Environmental Studies*, 14(4), 509-516.
- Shareef, H., Abbas, M. F., Jasim, A. M. (2022) Response of date palm offshoots (*Phoenix dactylifera* L.) to the foliar spray of salicylic acid and citric acid under salinity conditions. *Folia Oecologica*, 49(2), 130–136. <https://doi.org/10.2478/foecol-2022-0015>
- Shabani, N., Sayadi, M. H. (2012) Evaluation of heavy metals accumulation by two emergent macrophytes from the polluted soil: an experimental study. *Environmentalist*, 32, 91–98. <https://doi.org/10.1007/s10669-011-9376-z>
- Shahbazi, K., Beheshti, M. (2019) Comparison of three methods for measuring heavy metals in calcareous soils of Iran. *SN Appl. Sci.* 1, 1541. <https://doi.org/10.1007/s42452-019-1578-x>

- Shahzad, B., Tanveer, M., Rehman, A., Cheema, S. A., Fahad, S., Rehman, S., Sharma, A. (2018) Nickel; whether toxic or essential for plants and environment - A review. *Plant Physiology and Biochemistry*, 132, 641-651. <https://doi.org/10.1016/j.plaphy.2018.10.014>
- Shehata, S., Ismail, S., Zaghloul, A. (2019) Minimizing Hazards of Heavy Metals in Vegetable Farms using Phytoremediation Technique. *Alexandria Science Exchange Journal*, 40, 471-486. <https://doi.org/10.21608/asejaiqjsae.2019.50352>
- Shinta, Y. C., Zaman, B., Sumiyati, S. (2021) Citric Acid and EDTA as chelating agents in phytoremediation of heavy metal in polluted soil: a review. *IOP Conference Series Earth and Environmental Science*, 896(1), 012023. <https://doi.org/10.1088/1755-1315/896/1/012023>
- Sulaiman, F. R., Hamzah, H. A. (2018) Heavy metals accumulation in suburban roadside plants of a tropical area (Jengka, Malaysia). *Ecol Process*, 7, 28. <https://doi.org/10.1186/s13717-018-0139-3>
- Suman, J., Uhlik, O., Viktorova, J., Macek, T. (2018) Phytoextraction of Heavy Metals: A Promising Tool for Clean-Up of Polluted Environment? *Front Plant Sci.*, 9, 1476. <https://doi.org/10.3389/fpls.2018.01476>
- Tatu, G. L. A., Vladut, N. V., Voicea, I., Vanghele, N. A., Pruteanu, M. A. (2020) Removal of heavy metals from a contaminated soil using phytoremediation. *MATEC Web of Conferences*, 305, 00061. <https://doi.org/10.1051/mateconf/202030500061>
- Tayiem A.N., Fares O., Abu Lail B., Hamed O., Deghles A., BerishaA., JodehS., Azzaoui K., et al. (2025), Cellulose functionalized with amino and mercapto chelating groups for adsorbing Hg(II) from wastewater: Design, synthesis and theoretical studies, *Journal of Molecular Structure*, 1326, 141099, ISSN 0022-2860, <https://doi.org/10.1016/j.molstruc.2024.141099>
- Twajj, S. D., Al-Saedi, A. J. H., Kadem, Y. J., Fadhl, R. H. (2019) Effect of citric and malic acids concentration on some growth and yield parameters of *Vicia fabal* plant. *Plant Archives*, 19(2), 4062-4066.
- Twumasi, P., Tandoh M. A., Borbi M. A., Ajoke A. R., Owusu-Tenkorang E., Okoro R., Dumevi R. M. (2016) Assessment of the levels of cadmium and lead in soil and vegetable samples from selected dumpsites in the Kumasi Metropolis of Ghana. *African Journal of Agricultural Research*, 11(18), 1608-1616.
- Ullah, S., Liu, Q., Wang, S., Jan, A. U., Sharif, H. M. A., Ditta, A., Wang, G., Cheng, H. (2023) Sources, impacts, factors affecting Cr uptake in plants, and mechanisms behind phytoremediation of Cr-contaminated soils. *Science of The Total Environment*, 899, 165726. <https://doi.org/10.1016/j.scitotenv.2023.165726>
- Usman, K., Al-Ghouti, M. A., Abu-Dieyeh, M. H. (2019) The assessment of cadmium, chromium, copper, and nickel tolerance and bioaccumulation by shrub plant *Tetraenaqataranse*. *Sci Rep.*, 9(1), 5658. <https://doi.org/10.1038/s41598-019-42029-9>
- Vesselinov, V. V., Alexandrov, B. S., O'Malley, D. (2018) Contaminant source identification using semi-supervised machine learning. *Journal of Contaminant Hydrology*, 212, 134-142. <https://doi.org/10.1016/j.jconhyd.2017.11.002>
- Vincent, F., Manu, J. M., Chessed, G., Vandi, P., Francis, S., Norah, S. (2022) Assessment of Heavy Metals Content in Soil and Groundwater within the Vicinity of Automobile Mechanic Workshops in Yola and Jimeta Towns, Adamawa State, Nigeria. *J. Appl. Sci. Environ. Manage.*, 26(11), 1749- 1755.
- Waseem, M., Khilji, S. A., Tariq, S., Jamal, A., et al. (2024) Phytoremediation of heavy metals from industrially contaminated soil using sunflower (*Helianthus annus L.*) by inoculation of two indigenous bacteria. *Plant Stress*, 11, 100297. <https://doi.org/10.1016/j.stress.2023.100297>
- Waziri, M., Abdullahi, U., Audu, A. A., Kalimullah. (2016) Phytoremediation Potentials of Selected Plants in Industrially Contaminated Soils. *International Journal of Environmental Science and Development*, 7(10), 757 – 762. <https://doi.org/10.18178/ijesd.2016.7.10.875>
- Wieczorek, J., Baran, A., Bubak, A. (2023) Mobility, bioaccumulation in plants, and risk assessment of metals in soils. *Science of The Total Environment*, 882, 163574. <https://doi.org/10.1016/j.scitotenv.2023.163574>
- Wirosoedarmo, R., Anugroho, F., Hanggara, S. D., Gustinasari, K. (2018) Effect of Adding Chelating Agents on the Absorption of Zinc from Polluted Soil Sludge Textile Industrial Waste by Sunflower Plant (*Helianthus annuus L.*). *Applied and Environmental Soil Science*, 2018(3), 1 – 8. <https://doi.org/10.1155/2018/8259520>
- Woldeamanuale, T. B., Hassen, A. S. (2017) Toxicity Study of Heavy Metals Pollutants and Physico-Chemical Characterization of Effluents Collected from Different Paint Industries in Addis Ababa, Ethiopia. *J Forensic Sci & Criminal Inves.*, 5(5), 555685. <https://doi.org/10.19080/JFSCI.2017.05.555685>

- Xia, S., Song, Z., Jeyakumar, P., Bolan, N., Wang, H. (2020) Characteristics and applications of biochar for remediating Cr(VI)-contaminated soils and wastewater. *Environmental Geochemistry and Health*, 42(18), 1-25. <https://doi.org/10.1007/s10653-019-00445-w>
- Yan, A., Wang, Y., Tan, S. N., Mohd Yusof, M. L., Ghosh, S., Chen, Z. (2020) Phytoremediation: A Promising Approach for Revegetation of Heavy Metal-Polluted Land. *Front. Plant Sci.*, 11, 359. <https://doi.org/10.3389/fpls.2020.00359>
- Yang, Q., Yang, C., Yu, H., Zhao, Z., Bai, Z. (2021) The addition of degradable chelating agents enhances maize phytoremediation efficiency in Cd-contaminated soils. *Chemosphere*, 269, 129373. <https://doi.org/10.1016/j.chemosphere.2020.129373>
- Yang, Y., Jiang, M., Liao, J., Luo, Z., Gao, Y., Yu, W., He, R., Feng, S. (2022) Effects of Simultaneous Application of Double Chelating Agents to Pb-Contaminated Soil on the Phytoremediation Efficiency of *Indocalamusdecorus* Q. H. Dai and the Soil Environment. *Toxics*, 10, 713. <https://doi.org/10.3390/toxics10120713>
- Zakari, A., Audu, A. A. (2021) Evaluation of potentially toxic metals (ptms) accumulation and translocation by *Albizia lebbek* from industrial soil. *SWJ*, 16(2), 157 – 161.
- Zhou, P., Zeng, D., Wang, X., Tai, L., Zhou, W., Zhuoma, Q., Lin, F. (2022) Pollution Levels and Risk Assessment of Heavy Metals in the Soil of a Landfill Site: A Case Study in Lhasa, Tibet. *Int J Environ Res Public Health*, 19(17), 10704. <https://doi.org/10.3390/ijerph191710704>
- Zulkernain, N. H., Uvarajan, T., Ng, C. C. (2023) Roles and significance of chelating agents for potentially toxic elements (PTEs) phytoremediation in soil: A review. *Journal of Environmental Management*, 341, 117926. <https://doi.org/10.1016/j.jenvman.2023.117926>

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