



Assessment of metals (Cu, Ni) and metalloids (As) induced stress responses in Barley (*Hordeum vulgare*) and wheat (*Triticum aestivum*)

T. El Rasafi^{1*}, S. Bouda¹, M. Nouri² and A. Haddioui¹

¹ Laboratory of Biotechnology and Valorization of phylogenetic Resources, Faculty of Science and Techniques, University of Sultan Moulay Slimane, Beni Mellal, Morocco.

² Department of Biology, Faculty of polydisciplinary, University of Sultan Moulay Slimane, Morocco.

Received 22 October 2019,
Revised 21 April 2020,
Accepted 25 April 2020

Keywords

- ✓ Metals sensitivity,
- ✓ seedling growth,
- ✓ morphological parameters,
- ✓ plant tolerance

elrasafi_taoufik@hotmail.com
Phone: +212662 53 55 25;

Abstract

Soil contamination by metals and metalloids is known to treat human beings and cause a serious damage of the ecosystem and destroy the environment. The present study was performed to evaluate the effect of Cu, Ni and As stress on seed germination and early seedling growth of barley and wheat. The plant seeds were treated with 0, 10, 50, 100 and 200 mg L⁻¹ of the chosen elements. Based on the results, no effect was observed on seed germination whatever the metal used and the concentration applied (ranged between 96.6 and 100 % of barley and wheat final germination). Arsenic was found to decrease significantly root and shoot length, Tolerance index, root toxicity index and Seeds vigor index of seedlings of both plant species. By contrast, increasing Ni concentrations had a reduced effect on the parameter tested of barley seedlings. Results showed that barley was more sensitive to increasing levels of As and a high tolerance to Ni toxicity. Furthermore, the decreasing order effect of the tested elements on barley and wheat seedlings was As > Cu > Ni.

1. Introduction

Soil pollution by trace elements becomes a major world concern due to their harmful effects on human health, animals and plants and due to their high cleanup cost (billions of dollars) [1–3]. Trace elements are mostly realized into soil because of intensive human activities such as mining, industry, irrational use of pesticide and fertilizers [4–7]. Because of their persistence in the environment and non-degradability by chemicals or microbial activities [5,8], trace elements might enter to food chain [9] by direct contact or consumption of contaminated plants and animals [2,10,11] causing several diseases such as morphological abnormalities, physiological disorder and molecular perturbation [12–14]. Among trace elements, Cu and Ni are considered as essential micronutrient for plant growth and development but toxic at higher levels [15–18] while As is the most abundant and toxic metalloid in the environment [19,20]. Numerous works have reported the toxic effects of Cu, Ni and As on seed germination, early seedling growth and biomass of several plant species such as fenugreek, alfalfa, sorghum, mustard and wheat [7,20–23] in different growth conditions. In addition, plants in presence of excess of metals and metalloids might suffer from several symptoms such as germination inhibition, growth reduction, photosynthesis and water uptake disturbance, cellular damage, reactive oxygen species generation and biochemical and physiological disorder [7,24–28].

Due to the harmful effect of trace elements on environment, there has been a growing interest on ecological risk test for environmental bio-monitoring of trace elements [21]. Actually and due to the scientific research evolution, the interest was given to physiological, biochemical and molecular biomarkers such as enzyme activities, chlorophyll fluorescence, lipid peroxidation, genotoxicity and changes in the genomic DNA [21,24,29]. However,

testing the effect of trace elements in the level of seed germination and seedling growth is primordial and highly required [30] since seed germination is the initial step in the plant life cycle [28,31]. This test have been reported to be a rapid and simple test assess and evaluate metals toxicity [32–34] and considered as a suitable indicator of metals stress [7,35] because of the sensitivity of seeds at this stage of cycle life to the metals [25] and to the environmental changes conditions [36].

The objective of the present experiment was to test the impact of Cu, Ni and As exposure on germination and early seedling growth of barley (*Hordeum vulgare*) and wheat (*Triticum aestivum*). Seed germination, growth, dry biomass as well as seeds vigor index, tolerance index and root toxicity index were assayed under increasing concentrations of above elements. In developing countries like Morocco, mining activities and agriculture are important in the national economy, while they produce high amount of wastes highly contaminated with trace elements [37,38] causing a major environmental problems. Moreover, the selection of barley and wheat species for this test because these crops are highly cultivated in Morocco even at arid and semi-arid areas [39–41] and used mostly for feed of both animals and humans.

2. Material and Methods

2.1. Plant seeds and Metal(loids) treatments

Dry, healthy and uniform size seeds of barley (*Hordeum vulgare*) and wheat (*Triticum aestivum*) were surface sterilized by soaking them in 1% Sodium Hypochlorite solution for 10 min. The seeds were immersed during 1min in 70% Ethanol then rinsed with distilled water. This procedure was used to eliminate the possible fungi growth and bacterial contamination.

Increasing concentrations (0, 10, 50, 100 and 200 mg / L) of Cu, Ni and As were prepared by dissolving the metal elements Cu ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$), Ni ($\text{NiSO}_4 \cdot 6\text{H}_2\text{O}$) and As ($\text{Na}_2\text{HAsO}_4 \cdot 7\text{H}_2\text{O}$) in distilled water.

2.2. Germination experiment

The selected seeds were gently added to Petri dish covered with one layer of filter paper and previously imbibed with 5 ml of each treatment. Distilled water was used as a control. 3 replicates (Petri dishes) per treatment were prepared with 10 seeds of barley or wheat equally spaced (1cm) in each Petri dish. The dishes were then transferred to the oven for five days at temperature of $23 \pm 1^\circ\text{C}$.

2.3. Measurement of biological responses

Germination percentage (GP) was calculated by dividing the germinated seeds in each Petri dish over total seeds. Seeds were considered germinated when the radical reached at least 2 mm. Both root length (RL) and shoot elongation (SL) of both plant seedlings were measured using a ruler. Tolerance Index (TI), Root Toxicity Index (RTI) and Seeds Vigor Index were estimated using the formulas by Iqbal and Rahmati (1992), Wierzbicka et al. (2015) and Abdul-Baki and Anderson (1973), respectively:

$$\begin{aligned} \text{TI} &= (\text{RL}_{\text{treatment}} / \text{RL}_{\text{control}}) \times 100 \\ \text{RTI} &= ((\text{RL}_{\text{control}} - \text{RL}_{\text{treatment}}) / \text{RL}_{\text{control}}) \times 100 \\ \text{SVI} &= (\text{GP} \times \text{TPL}) / 100 \end{aligned}$$

With: GP = Germination percentage, $\text{RL}_{\text{control}}$ = Root length in control, $\text{RL}_{\text{treatment}}$ = Root length in treatment and TPL= Total plant length (TPL = roots + shoots length)

Production of dry biomass was calculated by drying seedlings in the oven for 48 h at 80°C .

2.4. Statistical analysis

In order to meet the normality assumptions of ANOVA, percentage data were arcsine-transformed. One-Way Analysis of Variance (ANOVA) followed by *dunnnett's post-hoc* test was applied to evaluate significant variations on the biological responses under metal treatments relatively to those observed under the respective controls ($p < 0.05$). The SPSS software (version 20.0) was used to perform the statistical analysis.

3. Results and discussion

3.1. Germination percentage

The obtained results of the present study showed that the stress caused by Cu, Ni and As had no significant effect on barley and wheat seed germination ($p > 0.05$) in comparison to control (Fig.1). In fact, whatever the metal used and the concentration applied both plant seeds reached almost 100% of final germination.

Germination assay is a rapid and easy test used to assess and evaluate metals phytotoxicity [32,44]. Phytotoxic effect of metals and metalloids on seeds germination is well documented. However, responses of plant seeds depends on the plant species, the metal itself and its concentration [20,27]. Several studies were carried out on barley [45–51] and wheat [21,26,52–56] seeds using both essential and non-essential elements under different experimental conditions. Sanal et al. (2014) found that both forms of arsenic (arsenate and arsenite) inhibited significantly germination seeds of barley. Different elements were found to have negative effects on barley such as Cu, Cd and Pb [46,58]. Cd was found to reduce germination percentage of wheat [59] while Cr had no negative effects on this parameter [60]. In soil, 10 plant species seeds (mung bean (*Phaseolus radiatus*), cucumber (*Cucumis sativus*), wheat (*Triticum aestivum*), sorghum (*Sorghum bicolor*), barley (*Hordeum vulgare*), Chinese cabbage (*Brassica campestris var. chinensis*), broccoli (*Brassica oleracea*), mustard (*Brassica nigra*), kale (*Brassica campestris*) and pea (*Pisum sativum*)) were exposed different forms of arsenic [20]. The authors found that germination of barley was not affected by arsenate (As^V) while wheat germination was negatively affected. Moreover, germination of wheat seeds was found reduced under Cr, Cd, Mn and Zn stress [56].

In our study, barley and wheat seeds showed a high tolerance to increasing levels of Cu, Ni and As and germinate even under the highest concentrations applied of the studied elements. This difference of findings might be due to concentration variation, cultivar differences and laboratory conditions.

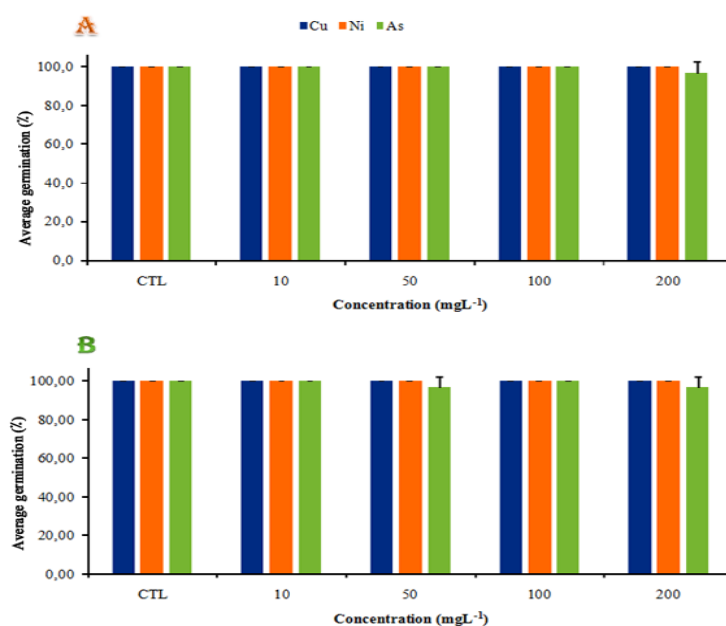


Fig.1. Effect of different concentrations of Cu, Ni and As on the average percentage (%) of germination of barley (A) and Wheat (B) grown in metal(loid)-spiked filter paper. Error bars represent standard deviation. Significant differences between test and reference (< reference) are indicated by “star*”. Statistical differences were tested at $p < 0.05$.

3.2. Root and shoot length

Increasing concentrations of the studied elements had different effects on barley and wheat root and shoot elongation according to the elements used and the dose applied (Fig. 2). Copper and arsenic reduced significantly both root and shoot length of barley seedlings compared to the control, with the most pronounced effect was observed when As levels was increased followed by Cu. Wheat root and shoot length showed the same pattern as that of barley. The most pronounced effect was found in presence of As especially in the highest concentration. For Ni, increasing concentrations have almost no effect on barley roots and wheat shoots (except in 200 mg L⁻¹

where the inhibition is significant). However, concentrations from 50 to 200 mg L⁻¹ reduced significantly both barley shoots and wheat roots. Mahmood et al. (2007) found that increasing Cu levels reduced significantly root length and a clear positive effect on shoot height of barley seedlings. In our experiment, As treatment decreased gradually the length of barley shoot which is in accordance with Sanal et al. (2014) that found that arsenate and arsenite reduced negatively both root and shoot of barley seedlings. The same findings were found when wheat (*Triticum aestivum*) were exposed to increasing As levels [54,62]. Root and shoots inhibition was found increased with increasing concentrations of As and Cd [55] as well as other elements such as Cd, B and Zn [63–65]. Arsenic is known to be toxic to several plant species from different families such as Fabaceae: *Trigonella foenum-graecum*, *Lathyrus sativus*, *Medicago sativa*, *Pisum sativum* [23,27,66] and Asteraceae: *Helianthus annuus* [67]. This effect of Arsenic might be due to their ability to be transported to plants via roots similarly to phosphate (Pi) using the same transporters [19,68] especially in *Holcuslanatus* and barley (*Hordeum vulgare*) plants [69]. Barley seedlings showed a high tolerance to Ni than Cu and As. It is worthy to note that low concentration of Cu (Cu₁₀ mg/L= 12.1 cm) and concentrations of Ni had a positive effect on barley and produced longer root (Ni₁₀ mg/L= 12.95 cm, Ni₅₀ mg/L= 12.54 cm and Ni₁₀₀ mg/L= 12.13 cm) comparing to the control (10.93cm). For wheat, only the concentration of 10 mg L⁻¹ of Cu and Ni and both 10 mg L⁻¹ and 50 mg L⁻¹ of Ni that increased the root length as compared to the control.

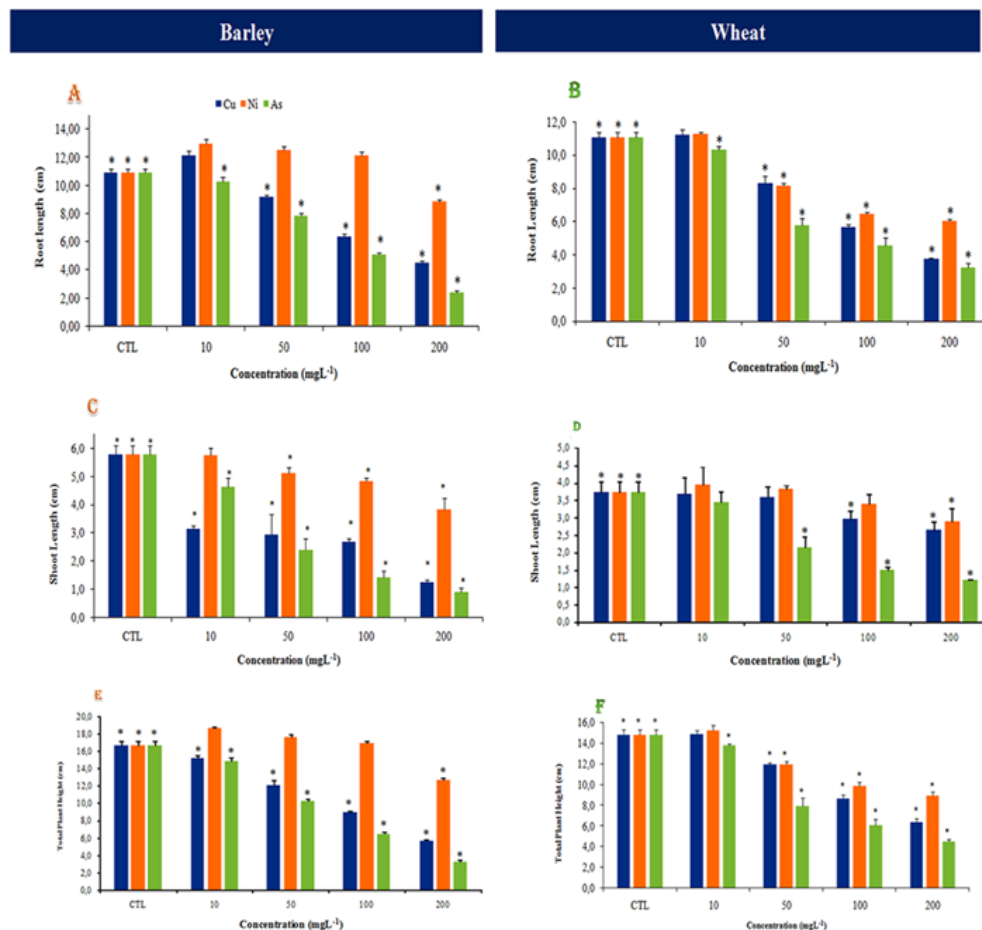


Fig.2. Effect of different concentrations of Cu, Ni and As on the root length, shoots length and total plant height of barley (A, C and E) and wheat (B, D and F) grown in metal (loid)-spiked filter paper. Error bars represent standard deviation. Significant differences between test and reference (< reference) are indicated by “star *”. Statistical differences were tested at p<0.05.

3.3. Tolerance index and root toxicity index

Roots developments of seedlings are very sensitive stage of plant growth and extremely sensitive to environmental changes [70] which make of them a rapid and useful way to study the mechanism of metal toxicity on plants [32,33,70]. In the present study, increasing Cu, Ni and As levels had different effects on root elongation of barley

and wheat seedlings which reflected by a difference of effects on tolerance index and root toxicity index (Fig. 3 and Fig. 4). Tolerance Index decreased significantly with increasing Cu and As concentrations, suggesting that these elements inhibited root elongation of barley and wheat. The same results were observed for wheat in presence of Ni while only the highest concentration of Ni (200 mg L⁻¹) that reduced significantly the TI of barley. Results indicated that high values of TI were recorded in presence of 10 mg L⁻¹ of Ni treatment (barley: 111.46%; wheat 102.1%) followed by Cu at the same concentration (barley: 110.85%; wheat: 101.26%). At the highest concentration (200 mg/L), tolerance degree of different elements was varied showing that the lowest TI value was recorded for As (22.18 % and 29.17 % for barley and wheat respectively) followed by Cu (41.26 % for barley and 33.79% for wheat). Ni was tolerated at all concentrations applied (at 200 mg L⁻¹, 81.10% and 54.47 % for barley and wheat respectively). In general, results suggest that barley seedlings were more tolerant to increasing concentrations of Ni than Cu and As. Mahmood et al. (2007) reported that increasing level of metals such as Cu induced a negative effect on tolerance index of barley. Likewise, other elements such as Cr, Cd, Pb, Mn, B and Zn were found to decrease the TI of wheat [26,56,61,63,64].

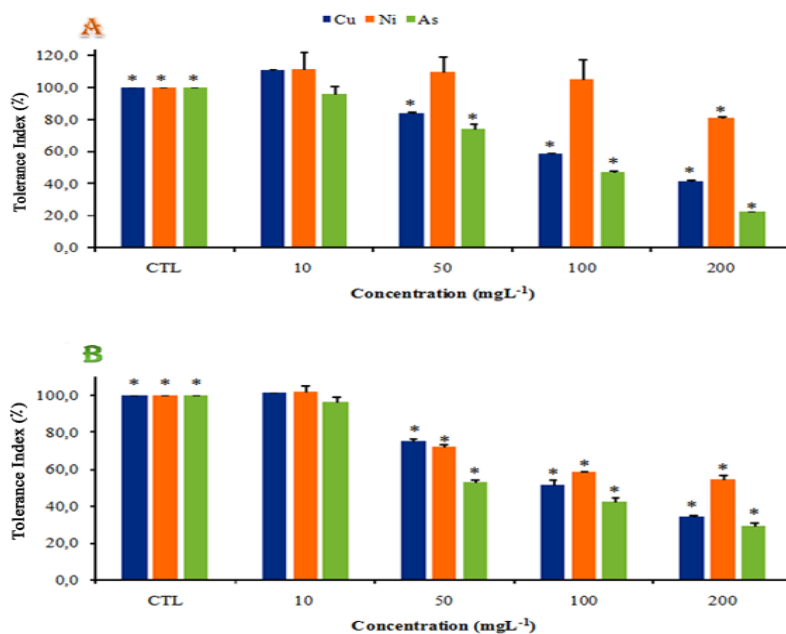


Fig.3. Effect of different concentrations of Cu, Ni and As on the Tolerance Index (%) computed for of barley (A) and wheat (B) grown in metal (loid)-spiked filter paper. Error bars represent standard deviation. Significant differences between test and reference (< reference) are indicated by “star *”. Statistical differences were tested at p<0.05.

Toxicity of As was higher than Cu and Ni toxicity with the increase of elements level. Higher values of RTI were recorded when As treatments were applied reaching almost 80% in the highest concentration (200 mg L⁻¹). Higher values of RTI indicated a higher toxicity of the substrate tested [42]. The results obtained indicated that As caused the most pronounced effect on root elongation than Cu for both species. However, increase of Ni treatments showed negative values of RTI at 10, 50 and 100 mg L⁻¹, while 19 % was recorded at 200 mg L⁻¹ (Fig. 4). For wheat, only 10 mg L⁻¹ of both Cu and Ni that induced a negative value of RTI while increased concentrations of the three elements tested showed an increase in RTI values. These results indicated low toxicity of Ni on barley seedlings even at high concentrations, followed by Cu than As in increasing tendency. Barley seedlings showed a high sensitivity to As stress than Cu or Ni.

3.4. Seeds vigor index

Figure 5 showed the results obtained for seeds vigor index of barley and wheat seeds in presence of increasing concentration of Cu, Ni and As. SVI was significantly reduced when As and Cu concentrations were increased starting from 10 mg L⁻¹ for barley and 50 mg L⁻¹ for wheat. Similarly, Ni reduced SVI value of wheat from 50 mg

L⁻¹ however, only the highest concentration of Ni (200 mg L⁻¹) that reduced significantly the SVI of barley. As compared to the control, As treatments induced the most pronounced effect on SVI leading to an extreme decrease at 200 mg L⁻¹. Arsenic was found to affect seedling development of barley [20,57], wheat [54,62] as well as other plant species such as mung bean [71] and sunflower [67]. Other metals such as Cr, Pb, B, Cu and Zn induced a decrease of seeds vigor index of other plant species (wheat, fenugreek, bean and tomato) [56,64,72–75]. As reported by Menon et al. (2016), the decrease of the SVI of barley might be due to decline in germination percentage and plant high under metals stress.

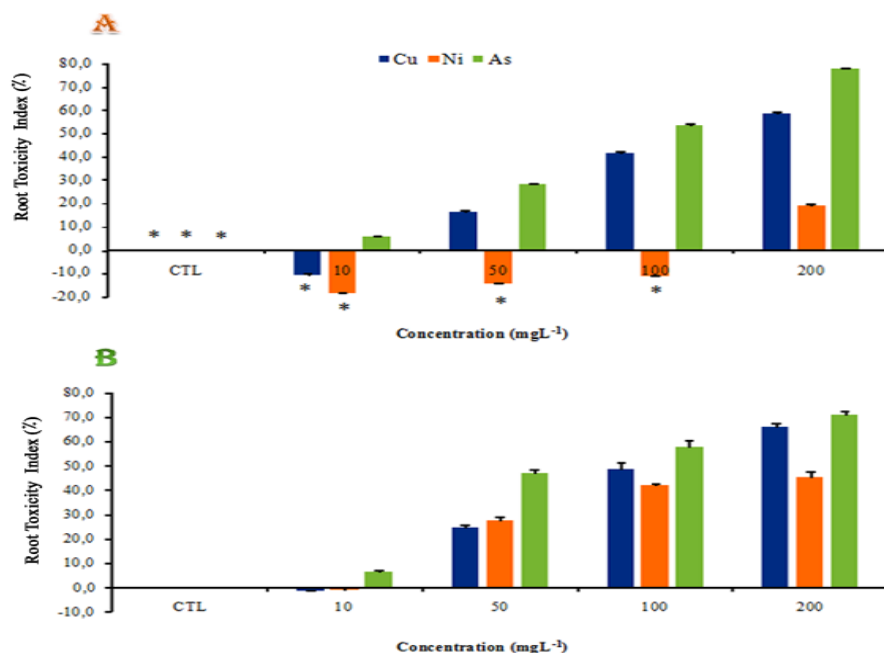


Fig.4. Effect of different concentrations of Cu, Ni and As on Root Toxicity Index computed for of barley (A) and wheat (B) grown in metal (loid)-spiked filter paper. Error bars represent standard deviation. Significant differences between test and reference (< reference) are indicated by “star*”. Statistical differences were tested at p<0.05.

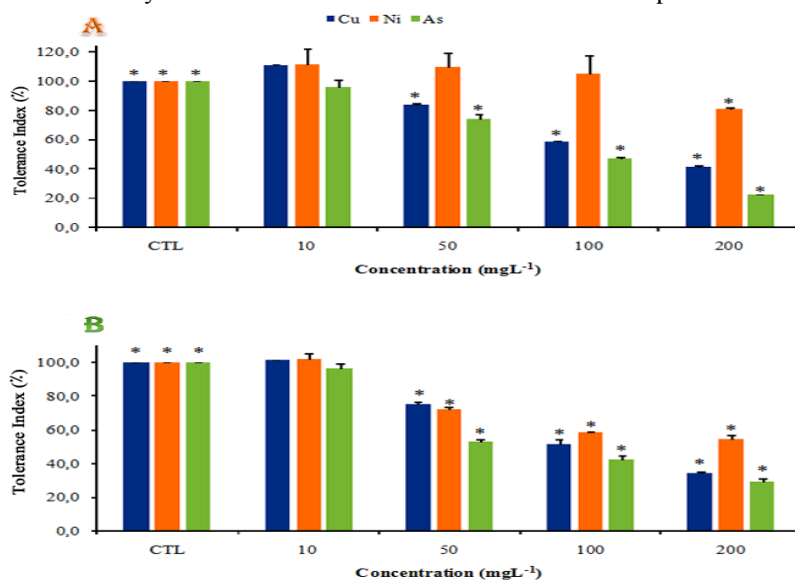


Fig.5. Effect of different concentrations of Cu, Ni and As on Seeds Vigor Index computed for of barley (A) and wheat (B) grown in metal (loid)-spiked filter paper. Error bars represent standard deviation. Significant differences between test and reference (< reference) are indicated by “star*”. Statistical differences were tested at p<0.05.

3.5. Dry biomass

In the present work, dry biomass of barley and wheat seedlings under different levels of Cu, Ni and As were higher than that found in the control in presence of all the concentrations tested (figure 6). Moreover, an increase

of dry biomass was recorded when the concentrations of all metals were increased. Comparing to the control (barley: 22.3 %), the highest dry biomass values were recorded at 200 mg L⁻¹ of Cu (46.3 %) and As (34.6 %) respectively. In contrast, As induced the highest value of dry biomass in wheat (39.66 %) followed by Ni (33.5 %) comparing to the control (20.55 %). Trace elements were found to modify several plant structure such as cell wall by affecting its porosity and plasticity and affecting the translocation of trace elements within xylem [17,76,77]. In addition, exposure of plants to Cu and As was found to enhance the biosynthesis of several organic compounds such cellulose, hemicellulose, pectin and lignin [76]. The cell wall rigidity and production of these compounds might be the reason of increasing plant biomass. Plants used this process as an adaptation strategy and mechanism of tolerance against numerous kind of stress such as metals stress [17,76,78,79].

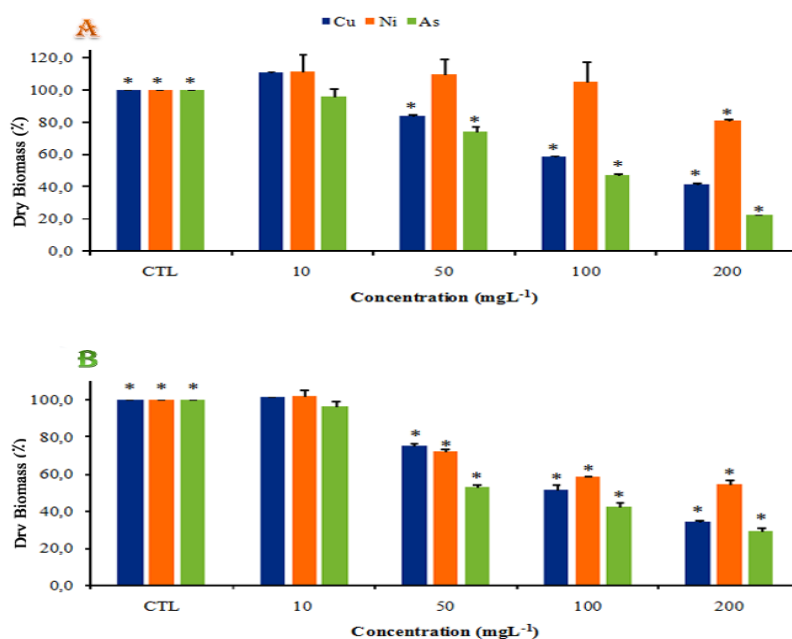


Fig.6. Effect of different concentrations of Cu, Ni and As on Dry Biomass computed for of barely (A) and wheat (B) grown in metal(loid)-spiked filter paper. Error bars represent standard deviation. Significant differences between test and reference (< reference) are indicated by “star *”. Statistical differences were tested at p<0.05.

4. Implications

In the present study, the response of barley and wheat to trace elements stress (Cu, Ni and As) during germination and early seedling growth was investigated. The results obtained revealed that the effect of these elements is extremely linked to the element itself and its concentration. Barley seeds were highly sensitive to increasing concentrations of As during seedling growth period. This has been reflected by a clear decrease of root and shoot length. Arsenic is easily taken up by root plants and translocated to aerial parts [80] using the same transporters as phosphate [68,81]. Once in plants, arsenic may generate reactive oxygen species which cause several biochemical and physiological damages in plants [19,80] as well as reduction of the plant morphological traits which is indicated by the present study.

Our results suggest that barley seeds showed a clear tolerance to increasing concentrations of Ni. The applied doses ranged from 10 to 200 mg L⁻¹ of Ni had almost no effect or reduced effect in the highest concentration on germination, root and shoot length, tolerance index, root toxicity index and dry biomass of barley seedling comparing to wheat. Ni have been reported as an essential elements [16,18] playing a major in plant life since it is a component of plant enzymes such as urease [16,82] and its deficiency may interrupt the plant life cycle [83]. The deficiency of Ni might affect the period of seeds maturation, earlier opening of seeds heads and several morphological symptoms (plants were less green, smaller) [83,84]. In this study, Cu was found to reduce the discussed parameters (except dry biomass) for both

plant species. Even that Cu is an essential element to plant life [15–17], but in excess it can be highly toxic and may induce several abnormalities in plants [15,75,85]. In general, the present results revealed that barley and wheat plant species showed a high sensibility to increasing levels of As. Furthermore, the general decreasing order effect of the tested elements on barley and wheat seedlings was As > Cu > Ni.

In this study, several questions were solved; nevertheless, others still remain un-discussed. We focused on early stage of seedling growth and studied the effect of single elements on germination and early seedling growth of the tested species. Further studies needed to be done in order to investigate the behavior of barley and wheat seeds in presence of a mixture of the tested elements since in soil a mixture of trace elements is always found. Moreover, this study should be extended for a long growing period in order to obtain enough plant biomass to analyze biochemical, physiological biomarkers such as MDA content, proline accumulations, enzymes activities, trace element translocation and accumulation in plant tissues. In addition, a molecular study is suitable to better understand the molecular response of both species to trace elements toxicity. The suggested analysis might help to increase our knowledge regarding trace element toxicity and better understand the mechanism of toxicity of Cu, Ni and As and plant strategies of tolerance/sensibility to excess of these elements.

References

1. C. Bopp, I. Christl, R. Schulin, M.W.H. Evangelou, Biochar as possible long-term soil amendment for phytostabilisation of TE-contaminated soils, *Environ. Sci. Pollut. Res.* 23 (2016) 17449–17458. <https://doi.org/10.1007/s11356-016-6935-3>.
2. H.S. Kim, K.R. Kim, H.J. Kim, J.H. Yoon, J.E. Yang, Y.S. Ok, G. Owens, K.H. Kim, Effect of biochar on heavy metal immobilization and uptake by lettuce (*Lactuca sativa L.*) in agricultural soil, *Environ. Earth Sci.* 74 (2015) 1249–1259. <https://doi.org/10.1007/s12665-015-4116-1>.
3. X. Wan, M. Lei, T. Chen, Cost–benefit calculation of phytoremediation technology for heavy-metal-contaminated soil, *Sci. Total Environ.* 563–564 (2016) 796–802. <https://doi.org/10.1016/j.scitotenv.2015.12.080>.
4. T. Abbas, M. Rizwan, S. Ali, M. Adrees, A. Mahmood, M. Zia-ur-Rehman, M. Ibrahim, M. Arshad, M.F. Qayyum, Biochar application increased the growth and yield and reduced cadmium in drought stressed wheat grown in an aged contaminated soil, *Ecotoxicol. Environ. Saf.* 148 (2018) 825–833. <https://doi.org/10.1016/j.ecoenv.2017.11.063>.
5. J. Luo, S. Qi, X.W.S. Gu, J. Wang, X. Xie, Evaluation of the phytoremediation effect and environmental risk in remediation processes under different cultivation systems, *J. Clean. Prod.* 119 (2016) 25–31. <https://doi.org/10.1016/j.jclepro.2016.01.043>.
6. P. Soudek, Š. Petrová, R. Vaňková, J. Song, T. Vaněk, Accumulation of heavy metals using Sorghum sp, *Chemosphere.* 104 (2014) 15–24. <https://doi.org/10.1016/j.chemosphere.2013.09.079>.
7. A. Elleuch, Z. Chaâbene, D.C. Grubb, N. Drira, H. Mejdoub, B. Khemakhem, Morphological and biochemical behavior of fenugreek (*Trigonella foenum-graecum*) under copper stress, *Ecotoxicol. Environ. Saf.* 98 (2013) 46–53. <https://doi.org/10.1016/j.ecoenv.2013.09.028>.
8. P. Xu, C.X. Sun, X.Z. Ye, W.D. Xiao, Q. Zhang, Q. Wang, The effect of biochar and crop straws on heavy metal bioavailability and plant accumulation in a Cd and Pb polluted soil, *Ecotoxicol. Environ. Saf.* 132 (2016) 94–100. <https://doi.org/10.1016/j.ecoenv.2016.05.031>.
9. T. Abbas, M. Rizwan, S. Ali, M. Zia-ur-Rehman, M. Farooq Qayyum, F. Abbas, F. Hannan, J. Rinklebe, Y. Sik Ok, Effect of biochar on cadmium bioavailability and uptake in wheat (*Triticum*

- aestivum* L.) grown in a soil with aged contamination, *Ecotoxicol. Environ. Saf.* 140 (2017) 37–47. <https://doi.org/10.1016/j.ecoenv.2017.02.028>.
10. P. Zhuang, M.B. McBride, H. Xia, N. Li, Z. Li, Health risk from heavy metals via consumption of food crops in the vicinity of Dabaoshan mine, South China, *Sci. Total Environ.* 407 (2009) 1551–1561. <https://doi.org/10.1016/j.scitotenv.2008.10.061>.
 11. J.H. Park, D. Lamb, P. Paneerselvam, G. Choppala, N. Bolan, J.W. Chung, Role of organic amendments on enhanced bioremediation of heavy metal(loid) contaminated soils, *J. Hazard. Mater.* 185 (2011) 549–574. <https://doi.org/10.1016/j.jhazmat.2010.09.082>.
 12. T. El Rasafi, M. Nouri, A. Haddioui, Metals in mine wastes: environmental pollution and soil remediation approaches – a review, *Geosystem Eng.* 9328 (2017) 1–16. <https://doi.org/10.1080/12269328.2017.1400474>.
 13. H. Ali, E. Khan, M.A. Sajad, Phytoremediation of heavy metals-Concepts and applications, *Chemosphere.* 91 (2013) 869–881. <https://doi.org/10.1016/j.chemosphere.2013.01.075>.
 14. M. Nouri, T. EL Rasafi, A. Haddioui, Levels Of Metals In Soils Of Ait Ammar Iron Mine , Morocco: Human Health Risks, *ACTA Chem. IASI.* 25 (2017) 127–144. <https://doi.org/10.1515/achi-2017-0011>.
 15. S.M.A. Dowidar, S.A. Abo-hamad, A.A. Mohsen, B.M.M. Khalaf, S.M.A. Dowidar, S.A. Abo-hamad, A.A. Mohsen, Bioremediation of copper stressed *Trigonella foenum graecum*, *Journal of Stress Physiology & Biochemistry* 9 (2013) 5–24.
 16. X.Y. Guo, Y.B. Zuo, B.R. Wang, J.M. Li, Y.B. Ma, Toxicity and accumulation of copper and nickel in maize plants cropped on calcareous and acidic field soils, *Plant Soil.* 333 (2010) 365–373. <https://doi.org/10.1007/s11104-010-0351-0>.
 17. M.B. Ali, N. Singh, A.M. Shohael, E.J. Hahn, K.Y. Paek, Phenolics metabolism and lignin synthesis in root suspension cultures of *Panax ginseng* in response to copper stress, *Plant Sci.* 171 (2006) 147–154. <https://doi.org/10.1016/j.plantsci.2006.03.005>.
 18. M. Yusuf, Q. Fariduddin, S. Hayat, A. Ahmad, Nickel: An overview of uptake, essentiality and toxicity in plants, *Bull. Environ. Contam. Toxicol.* 86 (2011) 1–17. <https://doi.org/10.1007/s00128-010-0171-1>.
 19. N. Garg, P. Singla, Arsenic toxicity in crop plants: Physiological effects and tolerance mechanisms, *Environ. Chem. Lett.* 9 (2011) 303–321. <https://doi.org/10.1007/s10311-011-0313-7>.
 20. Y. Yoon, W.M. Lee, Y.J. An, Phytotoxicity of arsenic compounds on crop plant seedlings, *Environ. Sci. Pollut. Res.* 22 (2015) 11047–11056. <https://doi.org/10.1007/s11356-015-4317-x>.
 21. K. Uruç Parlak, Effect of nickel on growth and biochemical characteristics of wheat (*Triticum aestivum* L.) seedlings, *NJAS - Wageningen J. Life Sci.* 76 (2016) 1–5. <https://doi.org/10.1016/j.njas.2012.07.001>.
 22. L. Kaur, K. Gardgil, S. Sharma, Assessment of Phytoextraction Potential of Fenugreek (*Trigonellafoenum-graecum* L.) to Remove Heavy Metals (Pb and Ni) from Contaminated Soil, *J. Chem. Heal. Risks.* 5 (2015) 1–14.
 23. C. Fulong, S. Wang, S. Mou, I. Azimuddin, D. Zhang, X. Pan, F.A. Al-Misned, M.G. Mortuza, Physiological responses and accumulation of heavy metals and arsenic of *Medicago sativa* L. growing on acidic copper mine tailings in arid lands, *J. Geochemical Explor.* 157 (2015) 27–35. <https://doi.org/10.1016/j.gexplo.2015.05.011>.
 24. I.A. Alaraidh, A.A. Alsahli, E.S. Abdel Razik, Alteration of antioxidant gene expression in response to heavy metal stress in *Trigonella foenum-graecum* L., *South African J. Bot.* 115 (2018) 90–93. <https://doi.org/10.1016/j.sajb.2018.01.012>.

25. C. Zayneb, K. Bassem, K. Zeineb, C.D. Grubb, D. Noureddine, M. Hafedh, E. Amine, Physiological responses of fenugreek seedlings and plants treated with cadmium, *Environ. Sci. Pollut. Res.* 22 (2015) 10679–10689. <https://doi.org/10.1007/s11356-015-4270-8>.
26. T. El Rasafi, M. Nouri, S. Bouda, A. Haddioui, The Effect of Cd, Zn and Fe on Seed Germination and Early Seedling Growth of Wheat and Bean, *Ekológia (Bratislava)*. 35 (2016) 213–223. <https://doi.org/10.1515/eko-2016-0017>.
27. A. Majeed, Z. Muhammad, S. Siyar, Assessment of heavy metal induced stress responses in pea (*Pisum sativum L.*), *Acta Ecol. Sin.* (2018) 12–16. <https://doi.org/10.1016/j.chnaes.2018.12.002>.
28. B. Márquez-García, C. Márquez, I. Sanjosé, F.J.J. Nieva, P. Rodríguez-Rubio, A.F. Muñoz-Rodríguez, The effects of heavy metals on germination and seedling characteristics in two halophyte species in Mediterranean marshes, *Mar. Pollut. Bull.* 70 (2013) 119–124. <https://doi.org/10.1016/j.marpolbul.2013.02.019>.
29. R. Banerjee, P. Goswami, K. Pathak, A. Mukherjee, Vetiver grass: An environment clean-up tool for heavy metal contaminated iron ore mine-soil, *Ecol. Eng.* 90 (2016) 25–34. <https://doi.org/10.1016/j.ecoleng.2016.01.027>.
30. X. Wang, C. Sun, S. Gao, L. Wang, H. Shuokui, Validation of germination rate and root elongation as indicator to assess phytotoxicity with *Cucumis sativus*, *Chemosphere*. 44 (2001) 1711–1721. [https://doi.org/10.1016/S0045-6535\(00\)00520-8](https://doi.org/10.1016/S0045-6535(00)00520-8).
31. M. Lamhamdi, A. Bakrim, A. Aarab, R. Lafont, F. Sayah, Lead phytotoxicity on wheat (*Triticum aestivum L.*) seed germination and seedlings growth, *Comptes Rendus - Biol.* 334 (2011) 118–126. <https://doi.org/10.1016/j.crvl.2010.12.006>.
32. S. Talebi, S.M.N. Kalat, A.L.S. Darban, The Study Effects of Heavy Metals on Germination Characteristics and Proline Content of Triticale (*Triticoseale Wittmack*), *Int. J. Farming Allied Sci.* 3 (2014) 1080–1087.
33. D. Lin, B. Xing, Phytotoxicity of nanoparticles: Inhibition of seed germination and root growth, *Environ. Pollut.* 150 (2007) 243–250. <https://doi.org/10.1016/j.envpol.2007.01.016>.
34. M. Di Salvatore, A.M. Carafa, G. Carratù, Assessment of heavy metals phytotoxicity using seed germination and root elongation tests: A comparison of two growth substrates, *Chemosphere*. 73 (2008) 1461–1464. <https://doi.org/10.1016/j.chemosphere.2008.07.061>.
35. O. Munzuroglu, H. Geckil, Effects of metals on seed germination, root elongation, and coleoptile and hypocotyl growth in *Triticum aestivum* and *Cucumis sativus*, *Arch. Environ. Contam. Toxicol.* 43 (2002) 203–213. <https://doi.org/10.1007/s00244-002-1116-4>.
36. R. Solanki, R. Dhankhar, Biochemical changes and adaptive strategies of plants under heavy metal stress, *Biologia (Bratisl)*. 66 (2011) 195–204. <https://doi.org/10.2478/s11756-011-0005-6>.
37. L. EL Founti, N. Saidi, A. Bouabdli, M. Saghi, M. Leblanc, Impact de la mine d'Aouli et de l'affluent Oued Za sur les sédiment et les matières en suspension de l'Oued Moulouya (Maroc), *Geomaghreb*. 1 (2003) 31–36.
38. A. Khalil, L. Hanich, R. Hakkou, M. Lepage, GIS-based environmental database for assessing the mine pollution : A case study of an abandoned mine site in Morocco, *J. Geochemical Explor.* 144 (2014) 468–477. <https://doi.org/10.1016/j.gexplo.2014.03.023>
39. A. Saidi, M. Diouri, A. Saidi, M. Diouri, G. Plan, Food self-sufficiency under the Green-Morocco Plan, *J. Exp. Biol. Agric. Sci.* 5 (2017) 33–40. <https://doi.org/halshs-01613992>.
40. L. Aziz, W. Sadok, Strategies used by the saffron producers of Taliouine (Morocco) to adapt to climate change, *Rev. Géographie Alp.* (2015) 0–13. <https://doi.org/10.4000/rga.2902>.
41. M. El Mourid, M. Karrout, Agriculture in arid and semi-arid regions of Morocco : Challenges and

- Prospects, *Rev. Al Awamia*. 92 (1996).
42. M. Wierzbicka, O. Bemowska-kałabun, B. Gworek, Multidimensional evaluation of soil pollution from railway tracks, *Ecotoxicology*. 24 (2015) 805–822. <https://doi.org/10.1007/s10646-015-1426-8>.
 43. A. Abdul-Baki, J.D. Anderson, Vigor Determination in Soybean Seed by Multiple Criteria, *Crop Sci*. 13 (1973) 630–633. <https://doi.org/10.2135/cropsci1973.0011183X001300060013x>.
 44. N. Kaur, T.E. Erickson, A.S. Ball, M.H. Ryan, A review of germination and early growth as a proxy for plant fitness under petrogenic contamination — knowledge gaps and recommendations, *Sci. Total Environ*. 603–604 (2017) 728–744. <https://doi.org/10.1016/j.scitotenv.2017.02.179>.
 45. R. Juknys, M. Račaite, G. Vitkauskaitė, J. Vencloviene, The effect of heavy metals on spring barley (*Hordeum vulgare* L.), *Zemdirbyste*, 96,2 (2009) 111–124.
 46. V. V. Talanova, A.F. Titov, N.P. Boeva, Effect of increasing concentrations of heavy metals on the growth of barley and wheat seedlings, *Russ. J. Plant Physiol*. 48 (2001) 100–103. <https://doi.org/10.1023/A:1009062901460>.
 47. J. Lachman, J. Dudjak, D. Miholová, D. Koliňová, V. Pivec, Effect of cadmium on flavonoid content in young barley (*Hordeum sativum* L.) plants, *Plant, Soil Environ*. 51 (2005) 513–516.
 48. H. Kudo, K. Kudo, M. Uemura, S. Kawai, Magnesium inhibits cadmium translocation from roots to shoots , rather than the uptake from roots , in barley, *Botany*. 93 (2015) 345–351. <https://doi.org/10.1139/cjb-2015-0002>.
 49. K. Kazuaki, H. Kudo, Y.K. Fujikawa, S. Kawai, The release of copper-induced phyto siderophores in barley plants is decreased by cadmium stress, *Botany*. 91 (2013) 568–572. <https://doi.org/10.1139/cjb-2013-0035>.
 50. M. Nouri, T.E.L. Rasafi, A. Haddioui, Responses of two barley subspecies to in vitro-induced heavy metal stress : seeds germination , seedlings growth and cytotoxicity assay, *Agric*. 65 (2019) 107–118. <https://doi.org/10.2478/agri-2019-0011>.
 51. Á. González, V. Chumillas, M.D.C. Lobo, Effect of Zn, Cd and Cr on growth, water status and chlorophyll content of barley plants (*H. vulgare* L.), *Agric. Sci*. 03 (2012) 572–581. <https://doi.org/10.4236/as.2012.34069>.
 52. M. Paunov, L. Koleva, A. Vassilev, J. Vangronsveld, V. Goltsev, Effects of different metals on photosynthesis: Cadmium and zinc affect chlorophyll fluorescence in durum wheat, *Int. J. Mol. Sci*. 19 (2018). <https://doi.org/10.3390/ijms19030787>.
 53. A. Humayan, K. Id, M. Rahman, U. Das, U. Sarkar, C. Roy, A. Reza, M.R. Talukder, A. Uddin, Reduction of cadmium toxicity in wheat through plasma technology, *PLOS ONE* (2019) 1–16. <https://doi.org/10.1371/journal.pone.0214509>
 54. X. Liu, S. Zhang, X. Shan, Y.G. Zhu, Toxicity of arsenate and arsenite on germination, seedling growth and amylolytic activity of wheat, *Chemosphere*. 61 (2005) 293–301. <https://doi.org/10.1016/j.chemosphere.2005.01.088>.
 55. X. Liu, S. Zhang, X. quan Shan, P. Christie, Combined toxicity of cadmium and arsenate to wheat seedlings and plant uptake and antioxidative enzyme responses to cadmium and arsenate co-contamination, *Ecotoxicol. Environ. Saf*. 68 (2007) 305–313. <https://doi.org/10.1016/j.ecoenv.2006.11.001>.
 56. I.R. Shaikh, P.R. Shaikh, R.A. Shaikh, A.A. Shaikh, Phytotoxic effects of Heavy metals (Cr, Cd, Mn and Zn) on Wheat (*Triticum aestivum* L.) Seed Germination and Seedlings growth in Black Cotton Soil of Nanded, India, *Res. J. Chem. Sci. Res. J. Chem. Sci*. 3 (2013) 2231–606.
 57. F. Sanal, G. Şeren, U. Guuner, Effects of arsenate and arsenite on germination and some

- physiological attributes of barley *hordeum vulgare* L., *Bull. Environ. Contam. Toxicol.* 92 (2014) 483–489. <https://doi.org/10.1007/s00128-014-1214-9>.
58. T. Kalai, K. Khamassi, J.A. Teixeira da Silva, H. Gouia, L. Bettaieb Ben-Kaab, Cadmium and copper stress affect seedling growth and enzymatic activities in germinating barley seeds, *Arch. Agron. Soil Sci.* 60 (2013) 765–783. <https://doi.org/10.1080/03650340.2013.838001>.
 59. M. de F.S. Guilherme, H.M. Oliveira, E. Da Silva, Cadmium toxicity on seed germination and seedling growth of wheat *Triticum aestivum*, *Acta Sci. Biol. Sci.* 37 (2015) 499. <https://doi.org/10.4025/acta sciobiolsci.v37i4.28148>.
 60. J.K. Datta, A. Bandhyopadhyay, A. Banerjee, N.K. Mondal, Phytotoxic effect of chromium on the germination, seedling growth of some wheat (*Triticum aestivum* L.) cultivars under laboratory condition., *J. Agric. Technol.* 7 (2011) 395–402.
 61. T. Mahmood, K.R. Islam, A.S. Muhammad, Toxic Effects of Heavy Metals on Early Growth and Tolerance of Cereal Crops, *Pak. J. Bot.* 39 (2007) 451–462.
 62. L.I. Chun-xi, F. Shu-li, S. Yun, J. Li-na, L.U. Xu-yang, H.O.U. Xiao-li, Effects of arsenic on seed germination and physiological activities of wheat seedlings, *J. Environ. Sci.* 19 (2007) 725–732.
 63. I. Ahmad, M.J. Akhtar, Z.A. Zahir, A. Jamil, I. Ahmad, E.T. Al, Effect of Cadmium on Seed Germination and Seedling Growth of Four Wheat (*Triticum Aestivum* L.) Cultivars, *Pak. J. Bot.* 44 (2012) 1569–1574.
 64. A. Habtamu, A.H. Ibrahim, F. Urgecha, N. Worku, Influence of boron on seed germination and seedling growth of wheat (*Triticum aestivum* L.), *African J. Plant Sci.* 8 (2014) 133–139. <https://doi.org/10.5897/AJPS2014.1148>.
 65. S.A. Moosavi, M.H. Gharineh, R. Tavakkol Afshari, A. Ebrahimi, Effects of Some Heavy Metals on Seed Germination Characteristics of Canola (*Barassica napus*), Wheat (*Triticum aestivum*) and Safflower (*Carthamus tinctorious*) to Evaluate Phytoremediation Potential of These Crops, *J. Agric. Sci.* 4 (2012) 11–19. <https://doi.org/10.5539/jas.v4n9p11>.
 66. D. Talukdar, Effect of Arsenic-induced Toxicity on Morphological Traits of *Trigonella foenum-graecum* L. and *Lathyrus sativus* L. During Germination and Early Seedling Growth, *Curr. Res. J. Biol. Sci.* 3 (2011) 116–123.
 67. M.A. Imran, M.N. Ch, R.M. Khan, Z. Ali, Toxicity of arsenic (As) on seed germination of sunflower (*Helianthus annuus* L.), *Int. J. Phys. Sci.* 8 (2013) 840–847. <https://doi.org/10.5897/IJPS2013.3894>.
 68. M.A. Farooq, F. Islam, B. Ali, U. Najeeb, B. Mao, R.A. Gill, G. Yan, K.H.M. Siddique, W. Zhou, Arsenic toxicity in plants: Cellular and molecular mechanisms of its transport and metabolism, *Environ. Exp. Bot.* 132 (2016) 42–52. <https://doi.org/10.1016/j.envexpbot.2016.08.004>.
 69. A.A. Meharg, M.R. Macnair, An altered phosphate uptake system in *Holcus lanatus* L., *New Phytol.* 116 (1990) 29–35. <https://doi.org/10.1111/j.1469-8137.1990.tb00507.x>
 70. Y.X. Chen, Y.F. He, Y.M. Luo, Y.L. Yu, Q. Lin, M.H. Wong, Physiological mechanism of plant roots exposed to cadmium, *Chemosphere.* 50 (2003) 789–793. [https://doi.org/10.1016/S0045-6535\(02\)00220-5](https://doi.org/10.1016/S0045-6535(02)00220-5).
 71. H. V Patel, S.R. Parmar, C.J. Chudasama, A. V Mangrola, Interactive studies of Zinc with Cadmium & Arsenic on seed germination and antioxidant properties of *Phaseolus aureus* Roxb, *Int. J. Plant, Anim. Environ. Sci.* 3 (2013) 166–174.
 72. P. Menon, N. Joshi, A. Joshi, Effect of Heavy Metals on Seed Germination of *Trigonella foenum-graceum* L, *Int. J. Life-Sciences Sci. Res.* 2 (2016) 488–493. doi.org/10.21276/ijlssr.2016.2.4.27.
 73. M. Mukherjee, Lead and Cadmium Toxicity on Seedling Growth and Metabolism of *Trigonella*

- foenum-graecum* L., *Int. J. Sci. Res.* 6 (2017) 1685–1689.
74. A. Habtamu, M. Shelema, R. Kedir, S. Ebsa, Seed germination and seedling growth of haricot bean (*Phaseolus vulgaris* L.) cultivars as influenced by copper sulphate, *World J. Agric. Sci.* 1 (2013) 312–317.
75. H. Ashagre, D. Almaw, T. Feyisa, Effect of copper and zinc on seed germination, phytotoxicity, tolerance and seedling vigor of tomato (*Lycopersicon esculentum* L. cultivar Roma VF), *Int. J. Agric. Sci. Res.* 2 (2013) 312–317. <http://academeresearchjournals.org/journal/ijasr>.
76. H. Le Gall, F. Philippe, J.-M. Domon, F. Gillet, J. Pelloux, C. Rayon, Cell Wall Metabolism in Response to Abiotic Stress, *Plants.* 4 (2015) 112–166. <https://doi.org/10.3390/plants4010112>.
77. D. da S. de Jesus, F.M. Martins, A.D. de Azevedo Neto, Structural changes in leaves and roots are anatomical markers of aluminum sensitivity in sunflower1, *Pesqui. Agropecuária Trop.* 46 (2017) 383–390. <https://doi.org/10.1590/1983-40632016v4641426>.
78. J.C.M.S. Moura, C.A.V. Bonine, J. de Oliveira Fernandes Viana, M.C. Dornelas, P. Mazzafera, Abiotic and biotic stresses and changes in the lignin content and composition in plants., *J. Integr. Plant Biol.* 52 (2010) 360–76. <https://doi.org/10.1111/j.1744-7909.2010.00892.x>.
79. N.H. Bhuiyan, G. Selvaraj, Y. Wei, J. King, Role of lignification in plant defense, *Plant Signal. Behav.* 4 (2009) 158–159. <https://doi.org/10.4161/psb.4.2.7688>.
80. P.M. Finnegan, W. Chen, Arsenic toxicity: The effects on plant metabolism, *Front. Physiol.* 3 JUN (2012) 1–18. <https://doi.org/10.3389/fphys.2012.00182>.
81. M.R. Shaibur, N. Kitajima, R. Sugawara, T. Kondo, S.M. Imamul Huq, S. Kawai, Effect of arsenic on phytosiderophores and mineral nutrition of barley seedlings grown in iron-depleted medium, *Soil Sci. Plant Nutr.* 55 (2009) 283–293. <https://doi.org/10.1111/j.1747-0765.2009.00360.x>.
82. H. Rahman, S. Sabreen, S. Alam, S. Kawai, Effects of nickel on growth and composition of metal micronutrients in barley plants grown in nutrient solution, *J. Plant Nutr.* 28 (2005) 393–404. <https://doi.org/10.1081/PLN-200049149>.
83. P.H. Brown, R.M. Welch, E.E. Cary, Nickel: A Micronutrient Essential for Higher Plants, *Plant Physiol.* 85 (1987) 801–803. <https://doi.org/10.1104/pp.85.3.801>.
84. P.H. Brown, R.M. Welch, E.E. Cary, R.T. Checkai, Beneficial effects of nickel on plant growth, *J. Plant Nutr.* 10 (1987) 2125–2135. <https://doi.org/10.1080/01904168709363763>.
85. G. Feigl, D. Kumar, N. Lehotai, N. Tugyi, Á. Molnár, A. Ördög, Á. Szepesi, K. Gémes, G. Laskay, L. Erdei, Z. Kolbert, Physiological and morphological responses of the root system of Indian mustard (*Brassica juncea* L. Czern.) and rapeseed (*Brassica napus* L.) to copper stress, *Ecotoxicol. Environ. Saf.* 94 (2013) 179–189. <https://doi.org/10.1016/j.ecoenv.2013.04.029>

(2020) ; <http://www.jmaterenvirosci.com>