



## Chronic exposure to Cerium Oxide Nanoparticles Induce Oxidative Stress in *Balb/c* Mice

F. Ndikuryayo<sup>1,2\*</sup>, Q. Zhong<sup>2</sup>, L. Xiong<sup>2</sup>, D. Liu<sup>2</sup>

<sup>1</sup>Centre Universitaire de Recherche et de Pédagogie Appliquées aux Sciences, Laboratoire de Nutrition-Phytochimie, d'Ecologie et Environnement Appliqués, Institut de Pédagogie Appliquée, Université du Burundi, BP 2700, Bujumbura, Burundi

<sup>2</sup>Hubei Key Laboratory of Genetic Regulation and Integrative Biology, College of Life Sciences, Central China Normal University, Wuhan 430079, Hubei, China.

\*Corresponding author, Email address: Ferdinand Ndikuryayo, email: [ferdinand.ndikuryayo@ub.edu.bi](mailto:ferdinand.ndikuryayo@ub.edu.bi)

Received 10 March 2021,  
Revised 13 July 2021,  
Accepted 15 July 2021

### Keywords

- ✓ Ceria nanoparticles
- ✓ *balb/c* mice
- ✓ Toxicity
- ✓ Oxidative stress

[ferdinand.ndikuryayo@ub.edu.bi](mailto:ferdinand.ndikuryayo@ub.edu.bi)  
Phone: +25769867706

### Abstract

Significant amounts of ceria nanoparticles (CeO<sub>2</sub> NPs) are released into the environment, whereas their effects on living organisms are not clearly established. We investigated the long-term toxicity of CeO<sub>2</sub> NPs to *Balb/c* Mice. Animals were exposed to 0, 100, 200, 400 and 800 mg/kg BW by intragastric administration (IGA) for 9 weeks. Weight was weekly recorded, whereas superoxide dismutase (SOD) activity and the levels of reduced glutathione (GSH) and malondialdehyde (MDA) were measured in livers and kidneys after exposure period. Although the weight of mice was not significantly affected, we observed a depletion of SOD and GSH contents at 400 and 800 mg/kg BW in the liver and kidney. Conversely, we noticed a substantial increase in MDA level in the liver at 400 and 800 mg/kg BW and in the kidney at 800 mg/kg BW. The results suggested that long-term exposure to CeO<sub>2</sub> NPs induces oxidative stress in mice, and hence highlighted the toxicity of these nanoparticles to mammals.

### 1. Introduction

Ceria nanoparticles (CeO<sub>2</sub> NPs) are used for various purposes, including catalyst, oxygen sensors, oxygen permeation membrane systems, fuel cells, and glass-polishing materials, and electrochromic thin-films [1]. Expected release of an important amount of CeO<sub>2</sub> NPs has raised health concerns [2] that the Organization for Economic Cooperation and Development (OECD) urged a rapid assessment of their toxicity [3]. This advice raised much attention of researchers who initiated a vast *ad hoc* campaign.

Results revealed a double face of CeO<sub>2</sub> NPs with regard to toxicity. On one hand, CeO<sub>2</sub> NPs could potentially prevent chronic inflammation and pathologies connected to oxidative stress [4-6]. This finding was confirmed in A549 and HepG2 cell lines shortly exposed to CeO<sub>2</sub> NPs although a long-term exposure of the same cell lines resulted in toxic effects [7]. Antioxidant effects were also noticed in mice shortly exposed to a lower and single dose of CeO<sub>2</sub> NPs [8]. On the other hand, cerium oxide (CeO<sub>2</sub>) was reported to cause pulmonary fibrosis in humans [9]. Recent studies highlighted the adverse effects caused by CeO<sub>2</sub> NPs to humans through the generation of reactive oxygen species (ROS) [5,10-14].

Using a single dose, Nemmar *et al.* [15] have recently observed oxidative stress in mice shortly exposed to CeO<sub>2</sub> NPs.

With regard to previous findings, the observed effects of CeO<sub>2</sub> NPs on either cell lines or mice are contradictory, and therefore deserve much attention. It is imperative to mention that most of these results were collected from *in vitro* experiments involving mammalian cells exposed to low and single doses of CeO<sub>2</sub> NPs. In addition, the dose-dependent chronic toxicity of these nanomaterials to mammals remains unknown. This study aimed at investigating the long-term effects of CeO<sub>2</sub> NPs to mice through the generation of oxidative stress.

## 2. Methodology

### 2.1 Chemical preparation

Ceria nanoparticles (<25 nm particle size and 30-50 m<sup>2</sup>/g Surface Area) were purchased from Sigma Aldrich (St. Louis, MO, USA). From a stock solution, five solutions (0, 10, 20, 40, 80 mg/mL) were prepared in normal sterilized saline, vortexed, and sonicated for 30 min to prevent particles from adhesion.

### 2.2 Animals treatment

*Balb/c* mice of 16-18 g body weight were bought from the Experimental Animal Center of Hubei, and grown according to the company's protocol. Mice were housed in stainless steel cages under controlled environmental conditions (temperature 21 ± 0.8 °C, humidity 50 ± 3% and a 12 h light/dark cycle). Commercial normal diet and water were freely available to animals. After acclimation for 1 week, 30 males were randomly separated into 5 groups of 6 mice each. Animals were then exposed to CeO<sub>2</sub> NPs through intragastric administration (IGA) to set final concentrations at 0, 100, 200, 400, and 800 mg/kg BW. Control group was exposed to normal saline.

### 2.3 Sample preparation

After 9-week exposure, mice were humanly killed. Livers and kidneys were rapidly collected, weighed, and homogenized in a physiological saline solution to get a 10% homogenate. Homogenates were centrifuged at 3000 rpm for 10 min at 4 °C (Eppendorf centrifuge 5415 R) and the supernatant was taken for analysis. The analyses were repeated three times and the data averaged.

### 2.4 Body weight and biochemical analysis

The body weight of mice was weekly recorded. Superoxide dismutase (SOD) activity was determined using a commercial kit (Nanjing Jiancheng Bioengineering Institute, Nanjing, China). Protein concentration was assayed according to the Bradford method [16]. Absorbance was measured using a microplate reader (BIO-TEC). SOD activity was expressed as units per mg of protein. Reduced glutathione (GSH) content was determined using a commercial kit (Nanjing Jiancheng Bioengineering Institute, Nanjing, China). Lipid peroxidation was ascertained as the content of malondialdehyde (MDA) generated by the thiobarbituric acid (TBA) reaction. 0.3 mL of sample or phosphate-buffered saline for the blank, 0.5 mL of 10% trichloroacetic acid (TCA) and 2 mL of 0.6% TBA prepared in 10% TCA were homogenized, heated in boiling water for 15 min and rapidly cooled on ice bath for 5 min. The mixture was centrifuged at 8400 rpm for 10 min and absorbance was measured at 450, 532 and 600 nm. MDA content was calculated using the Eqn.1.

$$MDA (\mu\text{mol/L}) = 6.45 (OD_{532} - OD_{600}) - 0.56 OD_{450}$$

Eqn. 1

## 2.5 Numerical and data analysis

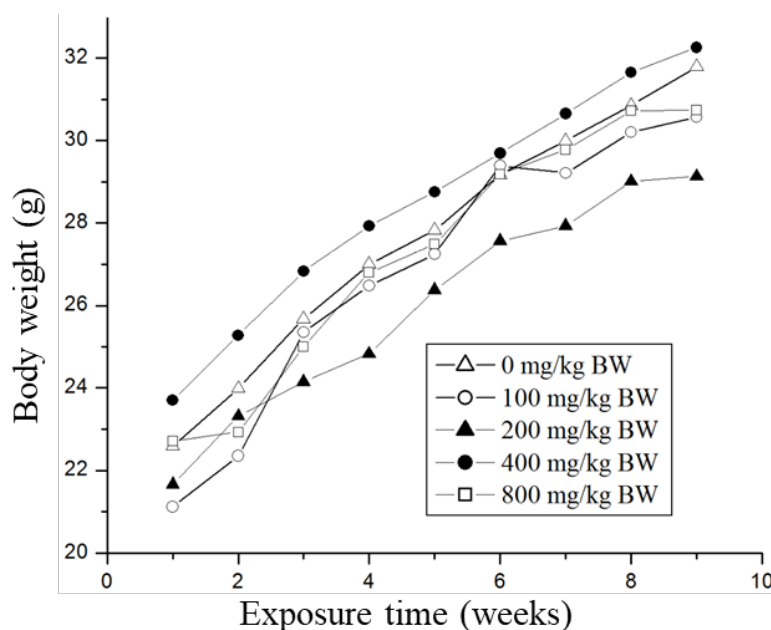
Microsoft Office Excel 2016 and Origin Pro 8 software were used for calculation and data analysis, respectively. One-Way Analysis of Variance (ANOVA) was undertaken to statistically analyze the significance between exposed and control groups. P-values less than 0.05 were considered to be significant.

## 3. Results and discussion

Hazardous health effects of nanoparticles are emerging issues. CeO<sub>2</sub> NPs are widely used [1] and, therefore, concerns about their potential toxicity to humans and environmental impact have increased quickly [3]. Nanoparticles can enter within cells, leading to oxidative stress that can attack almost any component of the cell [17]. Fortunately, cells possess antioxidant mechanisms that limit the harmful effects of oxidative stress. This defense system includes antioxidant molecules, including superoxide dismutase (SOD) and glutathione (GSH) commonly used as biomarkers of oxidative. In addition, animal weight and MDA content are also assessed for the same purpose.

### 3.1 Body weight

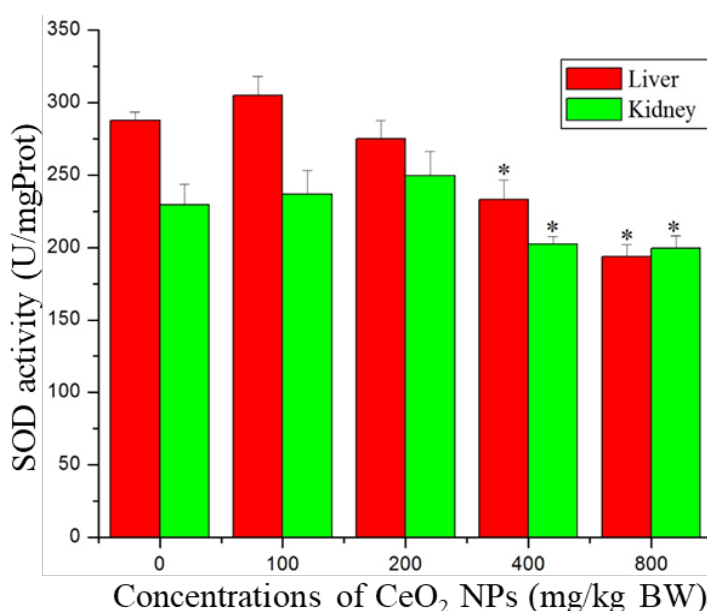
To follow their growth, we weekly recorded the body weight of mice for 9 weeks. As shown in **Figure 1**, there was no significant changes in body weight of experimental groups compared with the control group. This observation is presumably due to the toxicity that may occur at molecular level prior to organismal one [18].



**Figure 1.** Body weight of *balb/c* mice exposed to CeO<sub>2</sub> NPs

### 3.2 Superoxide dismutase activity

SOD is a critical enzyme involved in antioxidant defense by breaking down harmful oxygen molecules in cells [18]. After 9 week-exposure, the activity of SOD was assayed to examine the enzymatic defense against oxidative stress in mice exposed to various concentrations of CeO<sub>2</sub> NPs. As shown in **Figure 2**, a light but not significant increase in SOD was observed in both liver and kidney at 100 mg/kg BW. Increased SOD activity indicates the production of ROS [19,20] following the exposure of mice to CeO<sub>2</sub> NPs.



**Figure 2.** SOD activity in mice exposed to CeO<sub>2</sub> NPs for 9 weeks

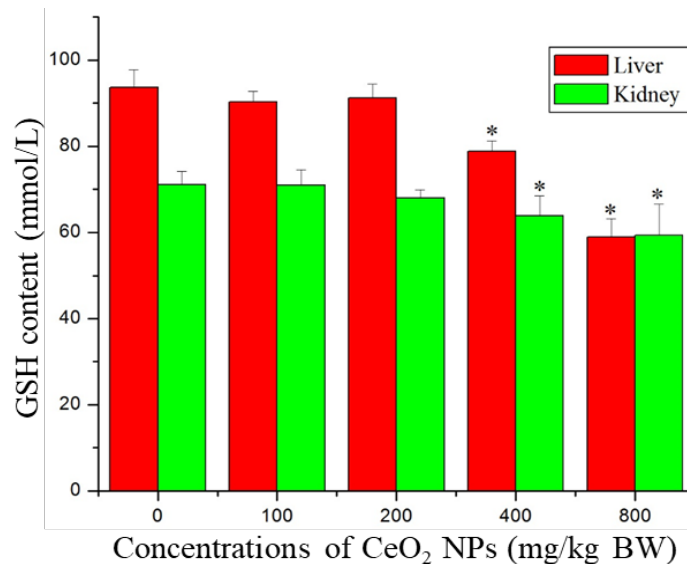
The observed increase in SOD activity is most probably due to hormesis that consists in activation of antioxidant systems, including SOD to overcome the ROS excess generated by the pollutant [20]. SOD activity continued to augment in kidney up to 200 mg/kg BW, suggesting that CeO<sub>2</sub> NPs toxicity was more important in liver than in kidney. On the other hand, SOD activity was significantly decreased in liver at 400 mg/kg BW ( $p < 0.01$ ) and 800 mg/kg BW ( $p < 0.01$ ) in concentration-dependent manner. Similar trend was also observed in kidney at the same concentrations, namely 400 mg/kg BW ( $p < 0.01$ ) and 800 mg/kg BW ( $p < 0.01$ ). Reduction in SOD activity has been found to be linked to oxidative stress [21]. These observations suggested not only the occurrence of oxidative stress but also the failure of the antioxidant system to fight against increased ROS. Although, this failure did not affect the animal's growth (Figure 1), animals could be affected after exposure to CeO<sub>2</sub> NPs for more long period.

### 3.3 GSH content

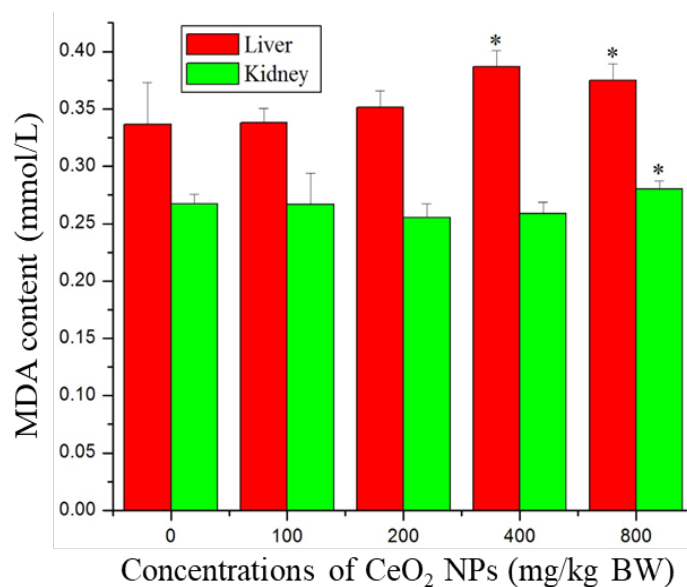
Found in all mammalian tissues, GSH is a tripeptide that prevents tissue damage by proving electron in peroxide detoxification. This small molecule protects mitochondria-rich tissues against oxidative stress [22]. It has been clearly established that GSH content decreases when oxidative stress increases. Figure 3 shows a significant depletion of GSH levels in liver at 400 mg/kg BW ( $p < 0.01$ ) and 800 mg/kg BW ( $p < 0.01$ ). This trend was noticed in kidney at the concentrations of 400 ( $p < 0.05$ ) and 800 mg/kg BW ( $p < 0.01$ ). We observed a higher amount of GSH in liver where it is mostly produced [23]. The augmentation of GSH suggested an adaptive reaction that prevents the damaging effects of ROS [18]. In accordance with SOD activity, continuous diminution of GSH level confirmed the failure of the antioxidant system to protect the animals. This trend was also observed in kidneys of mice exposed to silver nanoparticles through oral administration [24].

### 3.4 MDA content

MDA quantification is a common and facile method for the qualification of oxidative stress in biological materials. Increase in MDA level denotes the peroxidation of polyunsaturated fatty acids due to ROS, and that can lead to cell injury [18]. Thus, MDA level was ascertained to test the existence of oxidative stress in mice.



**Figure 3.** GSH content in mice exposed to CeO<sub>2</sub> NPs for 9 weeks



**Figure 4.** MDA content in mice exposed to CeO<sub>2</sub> NPs for 9 weeks

As depicted in **Figure 4**, MDA levels were significantly increased at 400 mg/kg BW and 800 mg/kg BW, and 800 mg/kg BW ( $p < 0.01$ ) in both liver and kidney. Increased levels of MDA have been reported to be related to oxidative stress in plants, humans and model systems [11]. In accordance with SOD activity and GSH content, these results clearly confirmed the occurrence of oxidative stress following the exposure of mice to CeO<sub>2</sub> NPs. Overall, the results suggested the occurrence of oxidative stress in mice exposed to CeO<sub>2</sub> NPs. With regard to the concentrations, the extent to which mice were affected was lower compared with the results reported by Nemmar *et al.* [15]. This discrepancy could be presumably associated with the particle size and the aggregation of CeO<sub>2</sub> NPs after administration.

## Conclusion

Mammalians, including humans, are exposed to CeO<sub>2</sub> NPs whose toxic effects are not clearly established. To investigate their toxicity, *Balb/c* mice were exposed to CeO<sub>2</sub> NPs, and changes in biomarkers of oxidative stress were recorded. We observed a significant depletion of SOD and GSH contents in the liver and kidney, and a substantial increase in MDA level in these organs. The results of

this study highlighted the *in vivo* toxicity of CeO<sub>2</sub> NPs to *balb/c* mice, and hence to humans, through the generation of oxidative stress following a long-term exposure. Therefore, people must be cautious during the handling or use of materials containing CeO<sub>2</sub> NPs. In the light of the growing use of CeO<sub>2</sub> NPs, additional investigations involving a very long period of exposure and offspring are highly required.

**Acknowledgement :** This work was supported by the Natural Science Foundation of China (30771429, 31071653), the Hubei Province Natural Science Foundation (2011CDB153) and the Fundamental Research Funds for the Central Universities Special (CCNU12A02001). The authors thank Professor Yang Xu for providing his laboratory used to grow *balb/c* mice and Désiré Nisubire for his input in the course of animal treatment.

**Disclosure statement:** *Conflict of Interest:* The authors declare that there are no conflicts of interest.

## References

- [1] A. Younis, D. Chu, S. Li, "Cerium oxide nanostructures and their applications," *Funct. Nanomater.* (2016) 53–68.
- [2] S.J. Snow, J. McGee, D.B. Miller, V. Bass, M.C. Schladweiler, R.F. Thomas, T. Krantz, C. King, A.D. Ledbetter, J. Richards, others, "Inhaled diesel emissions generated with cerium oxide nanoparticle fuel additive induce adverse pulmonary and systemic effects," *Toxicol. Sci.* 142 (2014) 403–417. <https://doi.org/10.1093/toxsci/kfu187>
- [3] OECD, "Environment directorate joint meeting of the chemical committee and the working party on chemicals," *Pesticides and Biotechnology*, (2008).
- [4] J. Niu, K. Wang, P.E. Kolattukudy, "Cerium oxide nanoparticles inhibits oxidative stress and nuclear factor- $\kappa$ B activation in H9c2 cardiomyocytes exposed to cigarette smoke extract," *J. Pharmacol. Exp. Ther.* 338 (2011) 53–61. <https://doi.org/10.1124/jpet.111.179978>
- [5] B.K. Pierscionek, Y. Li, R.A. Schachar, W. Chen, "The effect of high concentration and exposure duration of nanoceria on human lens epithelial cells", *Nanomedicine: NBM.* 8 (2012) 383–390. <https://doi.org/10.1016/j.nano.2011.06.016>
- [6] S.M. Hirst, A.S. Karakoti, R.D. Tyler, N. Sriranganathan, S. Seal, C.M. Reilly, "Anti-inflammatory properties of cerium oxide nanoparticles," *Small.* 5 (2009) 2848–2856. <https://doi.org/10.1002/sml.200901048>
- [7] L. De Marzi, A. Monaco, J. De Lapuente, D. Ramos, M. Borrás, M. Di Gioacchino, S. Santucci, A. Poma, "Cytotoxicity and genotoxicity of ceria nanoparticles on different cell lines *in vitro*," *Int. J. Mol. Sci.* 14 (2013) 3065–3077. <https://doi.org/10.3390/ijms14023065>
- [8] S.M. Hirst, A. Karakoti, S. Singh, W. Self, R. Tyler, S. Seal, C.M. Reilly, "Bio-distribution and *in vivo* antioxidant effects of cerium oxide nanoparticles in mice," *Environ. Toxicol.* 28 (2013) 107–118. <https://doi.org/10.1002/tox.20704>
- [9] J.W. McDonald, A.J. Ghio, C.E. Sheehan, P.F. Bernhardt, V.L. Roggli, "Rare earth (cerium oxide) pneumoconiosis: analytical scanning electron microscopy and literature review," *Mod. Pathol. an Off. J. United States Can. Acad. Pathol. Inc.* 8 (1995) 859–865.
- [10] E.-J. Park, J. Choi, Y.-K. Park, K. Park, "Oxidative stress induced by cerium oxide nanoparticles in cultured BEAS-2B cells," *Toxicology.* 245 (2008) 90–100. <https://doi.org/10.1016/j.tox.2007.12.022>
- [11] H.-J. Eom, J. Choi, "Oxidative stress of CeO<sub>2</sub> nanoparticles via *p38-Nrf-2* signaling pathway in human bronchial epithelial cell, *Beas-2B*," *Toxicol. Lett.* 187 (2009) 77–83. <https://doi.org/10.1016/j.toxlet.2009.01.028>



- [12] S. Hussain, F. Al-Nsour, A.B. Rice, J. Marshburn, B. Yingling, Z. Ji, J.I. Zink, N.J. Walker, S. Garantziotis, "Cerium dioxide nanoparticles induce apoptosis and autophagy in human peripheral blood monocytes," *ACS Nano*. 6 (2012) 5820–5829. <https://doi.org/10.1021/nn302235u>
- [13] G. Cheng, W. Guo, L. Han, E. Chen, L. Kong, L. Wang, W. Ai, N. Song, H. Li, H. Chen, "Cerium oxide nanoparticles induce cytotoxicity in human hepatoma *SMMC-7721* cells via oxidative stress and the activation of MAPK signaling pathways," *Toxicol. Vitro*. 27 (2013) 1082–1088. <https://doi.org/10.1016/j.tiv.2013.02.005>
- [14] A. Srinivas, P.J. Rao, G. Selvam, P.B. Murthy, P.N. Reddy, "Acute inhalation toxicity of cerium oxide nanoparticles in rats," *Toxicol. Lett.* 205 (2011) 105–115. <https://doi.org/10.1016/j.toxlet.2011.05.1027>
- [15] A. Nemmar, P. Yuvaraju, S. Beegam, M.A. Fahim, B.H. Ali, "Cerium oxide nanoparticles in lung acutely induce oxidative stress, inflammation, and DNA damage in various organs of mice," *Oxid. Med. Cell. Longev.* (2017). <https://doi.org/10.1155/2017/9639035>
- [16] M.M. Bradford, "A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding," *Anal. Biochem.* 72 (1976) 248–254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- [17] J.K. Fard, S. Jafari, M.A. Eghbal, "A review of molecular mechanisms involved in toxicity of nanoparticles," *Adv. Pharm. Bull.* 5 (2015) 447–454. <https://doi.org/10.15171/apb.2015.061>
- [18] V. Demidchik, "Mechanisms of oxidative stress in plants: from classical chemistry to cell biology," *Environ. Exp. Bot.* 109 (2015) 212–228. <https://doi.org/10.1016/j.envexpbot.2014.06.021>
- [19] A.M. Pisoschi, A. Pop, "The role of antioxidants in the chemistry of oxidative stress: A review," *Eur. J. Med. Chem.* 97 (2015) 55–74. <https://doi.org/10.1016/j.ejmech.2015.04.040>
- [20] R.A. Larson, The antioxidants of higher plants, *Phytochemistry*. 27 (1988) 969–978. [https://doi.org/10.1016/0031-9422\(88\)80254-1](https://doi.org/10.1016/0031-9422(88)80254-1)
- [21] X. Li, X. Ping, S. Xiumei, W. Zhenbin, X. Liqiang, "Toxicity of cypermethrin on growth, pigments, and superoxide dismutase of *Scenedesmus obliquus*," *Ecotoxicol. Environ. Saf.* 60 (2005) 188–192. <https://doi.org/10.1016/j.ecoenv.2004.01.012>
- [22] S.C. Lu, Glutathione synthesis, *Biochim. Biophys. Acta* (BBA) 1830 (2013) 3143–3153. <https://doi.org/10.1016/j.bbagen.2012.09.008>
- [23] M. Ookhtens, N. Kaplowitz, Role of the liver in interorgan homeostasis of glutathione and cysteine, *Semin Liver Dis*, 18 (1998) 313–329. <https://doi.org/10.1055/s-2007-1007167>
- [24] O.S. Adeyemi, I. Adewumi, T.O. Faniyan, "Silver nanoparticles influenced rat serum metabolites and tissue morphology," *J. Basic Clin. Physiol. Pharmacol.* 26 (2015) 355–361. <https://doi.org/10.1515/jbcpp-2013-0092>

(2021) ; <http://www.jmaterenvirosci.com>